



Research Journal of Pharmaceutical, Biological and Chemical Sciences

Antioxidant Activities of Some Thai and Exotic Fruits Cultivated in Thailand.

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ABSTRACT

The twenty-one methanolic extracts of meat or peel of native and exotic fruits cultivated in Thailand were determined their total phenolic content and antioxidant activities by Folin-Ciocalteu reagent and Trolox equivalent antioxidant capacity (TEAC) assay. The ripe fruit of *Carissa carandas* and the raw fruit of *Ficus hirta* presented the highest and lowest %yield of extract as 67.20% and 12.86%, respectively. The *Terminalia chebula* and *Baccaea motleyana* extracts contained the highest and lowest amount of total phenols, 6.96 and 0.05 g/100g dried fruit, respectively. The *T. chebula* and *Syzygium malaccense* extracts contained the highest and lowest amount of total phenols, 16.12 and 0.22 g/100g extract, respectively. The range of TEAC and IC₅₀ values of the fruit extracts were 0.35–0.003 and 21.33 µg/mL –5.10 mg/mL, respectively. The *T. chebula* and *Diospyros peregrina* extracts showed the highest antioxidant activities, while *B. motleyana* extracts indicated the lowest. The extracts of *T. Chebula* and *D. peregrina* (raw fruit) showed equal TEAC value, however the total phenolics content of *D. peregrina* raw fruit was quite low. Finally almost methanolic extracts of Thai and exotic fruits in this study presented their antioxidant activities in direct proportion to the amount of total phenolics.

Keywords: fruits, total phenolics, antioxidant activity, TEAC

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INTRODUCTION

Thailand has a lot of kinds of fruits that are different in shape, size, taste, smell and texture. Some of them are exotic fruits that have been cultivated in Thai territory for a long time. Most of the fruits are sweet and delicious and some are sour and astringent taste. They are sources of carbohydrate, vitamins, minerals and fiber. In Thailand, fruits are not only giving high nutritional values but the fruits and other parts of their trees also beneficial as traditional medicines as shown in Table 1 [1-6]. These pharmacological activities were interesting. Therefore several researchers have been investigated and reported useful scientific information about their activities as follow. The *Annona muricata* shows cytotoxic [7], antileishmanial [8], and molluscicides activities [9]. The *Averrhoa carambola* L indicates hypotensive [10], muscle relaxant [11], and HIV-1 reverse transcriptase inhibitors activities [12]. The *Baccaurea sapida* presents hypotensive activity [13]. The *Carissa carandas* Linn is mentioned histamine releasing [14] and cardiogenic activities [15]. The *Coccinia grandis* indicates hypoglycaemic [16], antiHIV-1 [17] and increasing movement of intestine activities [18]. The *Dillenia indica* is reported anticonvulsion, antidiabetes [19], antibacterial and antiviral activities [20]. The *Diospyros peregrina* has spermicidal [21], antiameobic, antiviral and hypoglycaemic activities [22]. The *Ficus hirta* presents cytotoxic and antiviral activities [23]. The *Ficus racemosa* has been reported on hypoglycaemic [24], hypotension [25], cytotoxic [26], antiviral [27], antibacterial [28], antipyretic and anti-inflammatory activities. The *Garcinia schomburgkiana* has anti-tumor promoting activity [29]. The *Lansium domesticum* indicates antimalarial [30-31], larvicidal [32], and insecticidal activities [33]. The *Mimusops elengi* shows diuretic [34], antiviral [35], antifungal [36] and spermicidal [37] activities. The *Terminalia chebula* has been reported on its antibacterial [38], antiviral [39], and antioxidant activities [40-41]. However there has no report about total phenolic contents and antioxidant activities of these fruits. The aim of this study is to determine the phenolic contents and antioxidant activities of these fruits.

Table 1 The common name, Thai name, scientific name, family and traditional used of fruits in this experiment.

| Scientific name | Common name | Family | Used for |
|--------------------------------------|----------------------------|---------------------------|--|
| <i>Angle Marmelos</i> | Bael, Bengal Quince, Bilak | Rutaceae | Antipyretic form malaria, flatulence (bark, root bark), influenza, bronchitis (watery from fresh leaf), drinking juice for tonic (slide of grill raw fruit), laxative (ripe fruit) |
| <i>Annona muricata</i> L. | Soursop | Annonaceae | Scurvy (ripe fruit), antiameobic (seed of raw fruit), vomit stimulant & haemostatic (seed), pesticide and poison to fish (seed) |
| <i>Ardisia elliptica</i> Thunb. S/ST | shoebutton | Myrsinaceae | Treat gonorrhoea (root) |
| <i>Averrhoa carambola</i> L. | Corambola, Star Fruit | Averrhoaceae, Oxalidaceae | Antipyretic (root, leaf), diarrhoea & antiameobic (stem bark), scurvy & laxative (fruit), Anthelmintic (flower) |
| <i>Baccaurea motleyana</i> | Lamai | Euphorbiaceae | Fruit can ate |
| <i>Baccaurea sapida</i> Muell. Arg. | Lotka | Euphorbiaceae | Vitamin C supplement, anti-thirst (fruit), antipyretic (all part), antidiarrhoea, anti-TB (root), anthelmintic & antifungal (leaf), |

| | | | |
|--|----------------------------------|--------------------------|---|
| <i>Carissa carandas</i> Linn | Karanda, Carunda, Christ's Thorn | Apocynaceae | Anthelmintic & appetite stimulant (root), diarrhoea & antipyretic & ear ach & sore throat (leaf), tonic(heartwood), scurvy & diarrhea (raw& ripe fruit) |
| <i>Coccinia grandis</i> Voigt | Ivy gourd | Cucurbitaceae | Vitamin A supplement (leaf), antipyretic & anti-diabetic (leaf, root), expectorant(leaf), anti-iching (leaf, flower), antivomitic (root), laxative (root bark), |
| <i>Dillenia indica</i> L. | Matat, chulta | Dilleniaceae | Mouth & throat astringent (leaf, bark), eat fruit, de-poisoning(root) |
| <i>Diospyros peregrina</i> Guerke | River ebony | Ebenaceae | Astringent & antipyretic & anti-amoebic (stem bark, raw fruit), heal lesions of oral mucosa, (bark, ripe fruit) antidiarrhoea (bark, raw fruit, seed) |
| <i>Ficus hirta</i> Vahl. | Hairy mountain fig | Moraceae | Antidiarrhoea, liver-,lung- and cardiotoxic (bark), |
| <i>Ficus racemosa</i> L. | Cluster fig, Country fig tree | Moraceae | Antidiarrhoea & anti-vomiting & wound healing (bark), antipyretic (root), flatulence (fruit) |
| <i>Garcinia duleis</i> Kurz | Mapood | Guttiferae | Expectorant, anticough, sore throat (fruit juice); astringent (bark); antipyretic (root) |
| <i>Garcinia schomburgkiana</i> Pierre | Madan | Clusiaceae, (Guttiferae) | Constipation, anti-cough, abnormal uterine bleeding |
| <i>Lansium domesticum</i> Correa <i>Aglaia dookko</i> Griff | Longkong | Meliaceae | Vitamin B & phosphorus supplement, heal lesions of oral mucosa, reduced body heat |
| <i>Lepisanthes fruticosa</i> (Roxb) Leenh. | Luna Nut | Sapindaceae | Antipyretic (root), antidiarrhoea (ripe fruit) |
| <i>Mimusops elengi</i> L. | Bullet wood | Sapotaceae | Gingival disorder (bark), cardiotoxic & antipyretic, headach (flower) |
| <i>Pouteria campechiana</i> Baehni | Canistel | Sapotaceae | Febrifuge, skin eruptions (bark); ulcers (seed) |
| <i>Spondias pinnata</i> (L.f.) Kurz | Hog plum | Anacardiaceae | Eat as vegetable (young shoot), vitamin C supplement (fruit), antithirst (fruit, root), diarrhoea (bark), anti- vomit, diuretic (bark) |
| <i>Syzygium malaccense</i> (L.) Merr. & L.M. Perry | Malay apple, Pomerac | Myrtaceae | Flatulent, antithirst |
| <i>Terminalia chebula</i> Retz | Myrobalan Wood | Combretaceae | Antipuretic, antiameobic antidiarrhoea, expectorant, antipyretic |

MATERIALS AND METHODS

Plant

The fruits were bought from local market or collected from orchard in Nakorn-Pathom province by the researchers of Faculty of Pharmacy, Silpakorn University in June, 2008. The voucher specimens were deposited in the Department of Pharmacognosy, Silpakorn University in Nakhon-Pathom, Thailand.

Chemicals

ABTS²⁻, 2,2'-azinobis-(3-ethylbenzthiazoline-6-sulfo-nate), was obtained as sulfonic acid from Sigma (St. Louis, USA). Trolox (or (+/-)-6-hydroxy-2,5,7,8-tetra-methyl-chroman-2-carboxylic acid) was purchased from Aldrich (Steinheim, Germany). Potassium persulfate, FeSO₄ x 7H₂O, and sodium acetate were purchased from Asia Pacific Specialty Chemicals Limited (Seven Hills, Australia). Folin-Ciocalteu reagent, FeCl₃ x 6H₂O and NaCl were purchased from Carlo ErbaReagenti (Milano, Italy). 2,4,6-tri-pyridyl-s-triazine (TPTZ) was obtained from Fluka Chemie GmbH (Switzerland) and methanol was purchased from Merck (Darmstadt, Germany).

Methanolic extract of fruits

The meat or peel of the fresh fruits were dried at 55 °C for 72 hrs before grinding and passing through sieve size 30 mesh. The methanolic extraction was performed by maceration of the dried samples in methanol in a ratio of sample to methanol of 1:4 for 72 hrs. The filtrates were evaporated using a rotary evaporator (Buechi R205, Switzerland). The dried extracts were kept at 4 °C until used.

Quantification of total polyphenols [42]

The 0.5 mL of methanolic fruit extracts (10 µg/mL) were mixed with 0.5 mL of Folin-Ciocalteu reagent and 0.5 mL 10% Na₂CO₃. The mixture was shaken and placed at room temperature for 1 hr before measuring the absorbance at 760 nm. The calibration curve was prepared using gallic acid with a concentration range of 2 - 8 mg/L. The total polyphenols was calculated and reported as gallic acid equivalent (GEA), g of gallic acid in 100 g of dried fruits and in 100 g of methanolic extracts.

Determination of antioxidant activity [43]

An antioxidant activity was performed by determination of scavenging effect on ABTS^{•+} radical. The ABTS^{•+} solution was prepared by mixing equal volume of 7 mM ABTS²⁻ in water with 4.9 mM potassium persulfate in water. The solution was protected from light and stored at room temperature for 12 – 16 hrs. ABTS^{•+} formation was checked for its absorbance (A) at 734 nm using UV-Vis-Spectrophotometer, (Agilent 8453E UV-Visible Spectroscopy System, Agilent Technology, USA.). The absorbance of ABTS^{•+} was equilibrated to 0.7 (± 0.02) by diluting with water at room temperature.

All samples were prepared in a concentration range of 0 - 10 µg/µL, except for that of *D. indica* (0-20 µg/µL), and of *C. carandas* and *A. elliptica* (0-50µg/µL). A portion of each dilution (50 µL) was mixed with 3 mL of ABTS^{•+} solution. After the mixture had been allowed to stand for 6 minutes at room temperature, its absorbance was measured at 734 nm using a spectrophotometer. Trolox was used as a standard. The antioxidant capacity was calculated as

an average of four replicate absorbance measurements, and reported as %inhibition along with IC_{50} and also as Trolox equivalent antioxidant capacity (TEAC).

Calculation of antioxidant capacity

Percent inhibition could be calculation as follows;

$$\% \text{ inhibition} = \frac{A (\text{solvent}) - A (\text{compound})}{A (\text{solvent})} \times 100$$

where A (solvent) was an absorbance of the sole solvent (no extract added) and A (compound) was an absorbance of the mixture of samples (or Trolox) and ABTS^{•+} solution at 734.

The curve was plotted between % inhibition and concentration of sample or Trolox solutions. The regression coefficient (r^2) was calculated from the linear curve. The IC_{50} was determined from the concentration that resulted in 50% inhibition. TEAC was the ratio of % inhibition of the sample to % inhibition of Trolox at the same concentration.

RESULT AND DISCUSSION

The %yield of methanolic extract of fruit in this experiment was shown in Table 2. The ripe fruit of *C. carandas* gave the highest %yield of extract (67.20%), while the raw fruit of *F. hirta* gave the lowest (12.86%). The ripe fruits of *A. elliptica*, *C. carandas*, *F. racemosa* and *M. elengi* gave higher amount of %yield of extract than their raw fruits, except that of *D. peregrine*. The amount of total phenols of dried fruits and crude extracts of most fruits in this experiment were rather low. The meat of *T. chebula* fruit had moderate % yield (43.17%), but contained the highest amount of total phenols, 6.96 ± 0.12 g/100 g of dried fruit and 16.12 ± 0.29 g/100 g extract. The *B. motleyana* contained the lowest amount of total phenols, 0.05 ± 0.00 g/100 g dried fruit, while the *S. malaccense* contained the lowest amount of total phenols, 0.22 ± 0.01 g/100g extract. The raw fruit of *D. peregrine* and *M. elengi* contained higher amount of total phenols in their dried fruit than that of their ripe fruits and the ripe fruit of *A. elliptica*, *C. carandas* and *D. peregrine* presented higher amount of total phenols in their crude extract than that of their raw fruits. Most of crude extracts and dried fruits in this experiment contained rather low amount of total phenols.

Table 2 The percent yield and amount of total-phenols (GAE) in dried fruits and extracts.

| Type of fruit extracts | Part of plant | %yield | Amount of total-phenols(GAE) | |
|--------------------------------------|-----------------|--------|-------------------------------------|---------------------------------|
| | | | g of gallic acid / 100g dried fruit | g of gallic acid / 100g extract |
| <i>Angle Marmelos</i> | fruit | 14.41 | 0.79 ± 0.02 | 5.51 ± 0.11 |
| <i>Annona muricata</i> L. | fruit | 50.86 | 0.65 ± 0.02 | 1.28 ± 0.04 |
| <i>Ardisia elliptica</i> Thunb. S/ST | raw fruit | 26.36 | 1.26 ± 0.05 | 3.04 ± 0.11 |
| | ripe fruit | 41.46 | 1.48 ± 0.10 | 5.64 ± 0.37 |
| <i>Averrhoa carambola</i> L. | fruit meat | 38.31 | 0.68 ± 0.01 | 1.77 ± 0.04 |
| <i>Baccaurea motleyana</i> | fruit | 18.51 | 0.05 ± 0.00 | 0.29 ± 0.01 |
| <i>Baccaurea sapida</i> Muell. Arg. | fruit meat | 59.36 | 0.22 ± 0.01 | 0.37 ± 0.02 |
| | fruit peel | 21.48 | 0.10 ± 0.02 | 0.47 ± 0.08 |
| | pacenta | 65.11 | 0.23 ± 0.01 | 0.71 ± 0.04 |
| <i>Carissa carandas</i> Linn | raw fruit | 44.66 | 0.24 ± 0.00 | 0.53 ± 0.01 |
| | ripe fruit | 67.20 | 0.82 ± 0.02 | 1.22 ± 0.02 |
| <i>Coccinia grandis</i> | ripe fruit | 25.31 | 0.23 ± 0.03 | 0.90 ± 0.14 |
| <i>Dillenia indica</i> L. | fruit | 32.95 | 0.80 ± 0.02 | 2.43 ± 0.07 |
| <i>Diospyros peregrina</i> Guerke | raw fruit | 35.580 | 3.34 ± 0.41 | 1.73 ± 1.16 |
| | ripe fruit | 19.76 | 1.07 ± 0.05 | 5.44 ± 0.24 |
| <i>Ficus hirta</i> Vahl. | fruit | 12.86 | 0.26 ± 0.01 | 1.98 ± 0.08 |
| <i>Ficus racemosa</i> L. | raw fruit | 13.61 | 0.20 ± 0.01 | 1.50 ± 0.08 |
| | ripe fruit | 18.85 | 0.21 ± 0.03 | 1.13 ± 0.15 |
| <i>Garcinia duleis</i> Kurz | fruit | 50.32 | 0.47 ± 0.05 | 0.94 ± 0.09 |
| <i>Garcinia schomburgkiana</i> | fruit meat | 42.13 | 0.56 ± 0.03 | 1.32 ± 0.08 |
| | leafless branch | 21.17 | 2.62 ± 0.22 | 12.37 ± 1.05 |
| <i>Lansium domesticum</i> Correa | fruit peel | 16.51 | 0.13 ± 0.03 | 0.81 ± 0.19 |
| <i>Lepisanthes fruticosa</i> (Roxb) | ripe fruit | 62.65 | 0.48 ± 0.02 | 0.77 ± 0.03 |
| <i>Mimusops elengi</i> L. | raw fruit | 32.97 | 4.58 ± 0.21 | 1.51 ± 0.07 |
| | ripe fruit | 38.61 | 3.26 ± 0.06 | 1.26 ± 0.02 |
| <i>Pouteria campechiana</i> Baehni | fruit | 31.93 | 0.16 ± 0.00 | 0.52 ± 0.01 |
| <i>Spondias pinnata</i> | fruit meat | 26.30 | 0.12 ± 0.01 | 0.46 ± 0.05 |
| | fruit peel | 28.74 | 0.31 ± 0.01 | 1.08 ± 0.03 |
| <i>Syzygium malaccense</i> (L.) | fruit | 42.92 | 0.09 ± 0.00 | 0.22 ± 0.01 |
| <i>Terminalia chebula</i> Retz | fruit meat | 43.17 | 6.96 ± 0.12 | 16.12 ± 0.29 |

 Table 3 The slope, intercept and r^2 of % inhibition equations, IC_{50} and TEAC values of the fruit extracts.

| Type of fruit extracts | Part of plant | Equations of methanol extract | | | IC_{50} ($\mu\text{g/mL}$) | TEAC |
|---------------------------|---------------|-------------------------------|-----------|--------|-----------------------------------|-------|
| | | slope ^a | intercept | r^2 | | |
| Trolox | | 5.0248 | -0.9708 | 0.9985 | 10.14 | 1 |
| <i>Angle Marmelos</i> | fruit | 0.0991 | 4.1147 | 0.9759 | 463.02 | 0.03 |
| <i>Annona muricata</i> L. | fruit | 0.0213 | 2.0219 | 0.8828 | 2252.49 | 0.008 |

| | | | | | | |
|---|-----------------|---------|---------|--------|---------|-------|
| <i>Ardisia elliptica</i> Thunb. S/ST | raw fruit | 0.0266 | 2.9073 | 0.9906 | 1770.40 | 0.01 |
| | ripe fruit | 0.3572 | 2.2732 | 0.9874 | 133.61 | 0.076 |
| <i>Averrhoa carambola</i> L. | fruit meat | 0.0515 | 1.1201 | 0.9659 | 949.12 | 0.012 |
| <i>Baccaurea motleyana</i> | fruit | 0.0097 | 0.6259 | 0.9110 | 5090.11 | 0.003 |
| <i>Baccaurea sapida</i> Muell. Arg. | fruit meat | 0.0164 | 1.2639 | 0.9272 | 2971.71 | 0.005 |
| | fruit peel | 0.0968 | 7.0874 | 0.9293 | 443.31 | 0.03 |
| | pacenta | 0.0584 | 0.9355 | 0.9955 | 840.15 | 0.013 |
| <i>Carissa carandas</i> Linn | raw fruit | 0.0219* | 12.4910 | 0.8444 | _* | _* |
| | ripe fruit | 0.3622 | 1.6861 | 0.9940 | 133.39 | 0.08 |
| <i>Coccinia grandis</i> Voigt | ripe fruit | 0.0369 | 1.1844 | 0.9780 | 1322.92 | 0.01 |
| <i>Dillenia indica</i> L. | fruit | 0.0748 | 5.0707 | 0.9825 | 600.66 | 0.025 |
| <i>Diospyros peregrina</i> Guerke | raw fruit | 1.6991 | 3.1351 | 0.9933 | 27.58 | 0.35 |
| | ripe fruit | 0.7468 | 2.6712 | 0.9937 | 63.38 | 0.15 |
| <i>Ficus hirta</i> Vahl. | raw fruit | 0.1048 | 0.7298 | 0.9973 | 470.14 | 0.02 |
| <i>Ficus racemosa</i> L. | raw fruit | 0.0506 | 0.0890 | 0.9758 | 986.38 | 0.01 |
| | ripe fruit | 0.0429 | 0.7464 | 0.9941 | 1148.10 | 0.01 |
| <i>Garcinia schomburgkiana</i> | fruit meat | 0.0701 | 3.8099 | 0.9430 | 658.92 | 0.02 |
| | leafless branch | 1.2576 | 4.0073 | 0.9874 | 36.57 | 0.26 |
| <i>Garcinia duleis</i> Kurz | fruit | 0.0629 | 3.1221 | 0.9614 | 745.28 | 0.02 |
| <i>Lansium domesticum</i> Correa | fruit peel | 0.0462 | 1.5003 | 0.9869 | 1049.78 | 0.012 |
| <i>Lepisanthes fruticosa</i> (Roxb) | ripe fruit | 0.0711 | 1.0821 | 0.9967 | 688.02 | 0.02 |
| <i>Mimusops elengi</i> L. | raw fruit | 1.2009 | 7.7204 | 0.9462 | 35.21 | 0.26 |
| | ripe fruit | 0.2810 | 3.6359 | 0.9841 | 165.00 | 0.06 |
| <i>Pouteria campechiana</i> Baehni | fruit | 0.0198 | 1.1197 | 0.9526 | 2468.70 | 0.006 |
| <i>Spondias pinnata</i> (L.f.) Kurz | fruit meat | 0.0131 | 0.6237 | 0.9610 | 3769.18 | 0.004 |

| | | | | | | |
|---------------------------------|------------|--------|--------|--------|---------|-------|
| | fruit peel | 0.0532 | 1.4589 | 0.9904 | 912.43 | 0.014 |
| Syzygium malaccense (L.) | fruit | 0.0186 | 0.5595 | 0.9830 | 2658.09 | 0.005 |
| Terminalia chebula Retz | fruit meat | 1.7182 | 5.3856 | 0.9825 | 25.97 | 0.35 |

The antioxidant of all extracts of fruits in this experiment was rather low as shown in Table 3. The highest antioxidant activity (TEAC = 0.35) were shown in *T. Chebula* and *D. peregrine*. The *T. Chebula* indicated the highest amount of total phenolics, and antioxidant activity. These results were according to the report of Bajpai. *et.al.*, 2005 that *T. Chebula* fruit was a source of gallic acid and had good antioxidant activities [44]. The extract that showed inverse proportion between its amount of total phenolics contents and its TEAC values was *D. peregrine*. The TEAC of *D. peregrine* raw fruit extract was 0.35 equal to *T. Chebula*. However, the total phenolics content in *D. peregrine* extract was 1.73 g/100 g crude extract comparing to 16.12 g/100 g crude extract of *T. Chebula*. The other inverse result was the total phenolic content of *D. peregrine* raw fruit that was lower than ripe fruit, but the raw fruit presented higher TEAC value. For comparing between fruit meat and leafless branch of *G. schomburgkiana*, the total phenolic content of the leafless branch was not only higher than the fruit meat, but also the antioxidant activity. For *B. motleyana* fruit extract, it gave the lowest antioxidant activity which according to low amount of total phenolic content in its extract. In addition, the *S. malaccense* extract presented the lowest total phenolic content which related to its low antioxidant activity. In this study the *A. carambola* fruit extract contained rather low antioxidant activity (TEAC = 0.012). However Shui and Leong reported that *A. carambola* fruit juice was good source of antioxidant activities, which its antioxidant activities were attributed to L-ascorbic, (-)-epicatechin and gallic acid in gallotannin forms [45].

CONCLUSION

The total phenolic contents and antioxidant activities of fruit extracts in this experiment were rather low. The *T. Chebula* fruit meat extracts showed highest total phenolic content and antioxidant activity. Most fruits, that more than one type of crude fruit of each fruit (eg. meat, peel, raw fruit and ripe fruit) was extracted, indicated their antioxidant activities direct proportion to their amount of total phenols. The correlation between amount of total phenolics and antioxidant activities in raw and ripe fruit could not make a conclusion. However, if the raw fruit presented higher antioxidant activity than the ripe fruit, it showed distinctive higher. But when the ripe fruit gave higher antioxidant activity than the raw fruit, it showed small higher, except *F. ramosa* that raw and ripe fruit had equal antioxidant activities. The rather low antioxidant activities of most extracts in this study may correlate to the low amount of total phenolic content in the extracts.



ACKNOWLEDGEMENT

This research work was kindly supported by Natural Research Council of Thailand, NRCT. We would like to thanks the Faculty of Pharmacy, Silpakorn University and Associated Prof. Dr. Sindhchai Keokitichai for the facilities and beneficially advise.

REFERENCES

- [1] Bunyaphatsara N, Chokchajareonporn A. Sa-mund-phai Mai-pan-ban. No.2. Prachachon Co., Bangkok. 1998.
- [2] Bunyaphatsara N, Chokchajareonporn A. Sa-mund-phai Mai-pan-ban. No.3. Prachachon Co., Bangkok. 1999.
- [3] Bunyaphatsara N, Chokchajareonporn A. Sa-mund-phai Mai-pan-ban. No.4. Prachachon Co., Bangkok. 2000.
- [4] Na Songka, B. Sa-mund-phai Thai No.1 second ed., Funny Publishing Ltd., Bangkok. Pahonyothin, Bangkok. 1982.
- [5] Poopatpong L. Sa-mund-phai Thai No.3 Funny Publishing Ltd., Pahonyothin, Bangkok. 1982.
- [6] Poopatpong L. Sa-mund-phai Thai No.5 Chutima Kaprim Ltd., Phayathai, Bangkok. 1987
- [7] Leaman DJ, Arnason JT, Yusuf R, Sangat-Roemantyo H, Soedjito H, Angerhofer CK, Pezzuto JM. J Ethnopharmacol 1995; 49: 1-16.
- [8] Jaramillo MC, Arango GJ, Gonzalez MC, Robledo SM, Velez ID. Fitoterapia 2000; 71(2):183-186.
- [9] Luna Jde S, De Carvalho JM, De Lima MR, Bieber LW, Bento Ede S, Franck X, Sant'ana AE. Nat Prod Res 2006; 20(3):253-257.
- [10] Padmawinata K, Hoyaranda E. Abstract of the 4th Asian Symp Med Plants Spices, Bangkok, Thailand. 1980; 159.
- [11] Mookhasmit M, Ngarmwathana W, Sawasdimongkol K, Permiphat U. J Med Ass Thailand 1971; 54(7):490-504.
- [12] Tan GT, Pezzuto JM, Kinghorn AD, Hughes SH. J Nat Prod 1991; 54(1):143-154.
- [13] Dhar ML, Dhar MN, Dhawan BN, Mehrotra BN, Srimal RC, Tandon JS. Indian J Exp Biol 1973; 11:43-54.
- [14] Joglekar SN, Gaitonde BB. Jpn J Pharmacol 1970; 20(3):367-372.
- [15] Vohra MM, DE NN. Indian J Med Res 1963; 51:937-940.
- [16] Kumar GP, Sudheesh S, Vijayalakshmi NR. Planta Med 1993; 59(4):330-332.
- [17] Tan GT, Pezzuto JM, Kinghorn AD, Hughes SH. J Nat Prod 1991; 54(1):143-154.
- [18] Sookvanichsilp N, Silpa-Archa W, Laemongkol D, Tanawiriyakul P, Tongkowi H. Asian J Pharm Suppl 1986; 6(8):132.
- [19] Bhakuni OS, Dhar ML, Dhar MM, Dhawan BN, Mehrotra BN. Indian J Exp Biol 1969; 7:250-262.
- [20] Uppalapati L, Rao JT. Chem Petro Chem J 1979; 10(8):21-23.
- [21] Choudhary DN, Singh JN, Verma SK, Singh BP. Indian J Exp Biol 1990; 28(8):714-716.
- [22] Dhar ML, Dhar MM, Dhawan BN, Mehrotra BN, Ray C. Indian J Exp Biol 1968; 6:232-247.

- [23] Bhakuni DS, Goel AK, Jain S, Mehrotra BN, Pat naik GK, Prakash V. Indian J Exp Biol 1988; 26(11):883-904.
- [24] Jain SR, Sharma SN. Planta Med 1967; 15(4):439-442.
- [25] Bharkuni DS, Dhar ML, Dhar MM, Dhawan BN, Gupta B, Srimali RC. Indian J exp Biol 1971; 9:91.
- [26] Bhakuni DS, Goel AK, Jain S, Mehrotra BN, Patnaik GK, Prakash V. Indian J Exp Biol 1988; 26(11):883-904.
- [27] Singh R. Phytopathol Mediterr 1971; 10:211.
- [28] Forestieri AM, Pizzimenti FC, Monforte MT, Bisignano G. Pharmacol Res Commun Suppl 1988; 20(5):33-36.
- [29] Murakami A, Jiwajiinda S, Koshimizu K, Ohigashi H. Cancer Lett 1995; 95 (1/2):137-146.
- [30] Yapp DT, Yap SY. J Ethnopharmacol 2003; 85(1):145-150.
- [31] Omar S, Zhang J, MacKinnon S, Leaman D, Durst T, Philogene BJ, Arnason JT, Sanchez-Vindas PE, Poveda L, Tamez PA, Pezzuto JM. Curr Top Med Chem 2003; 3(2):133-139.
- [32] Monzon RB, Alvir JP, Luczon LL, Morales AS, Mutuc FE. Southeast Asian J Trop Med Public Health 1994; 25(4):755-759.
- [33] Leatemala JA, Isman MB. Phytoparasitica 2004; 32(1):30-37.
- [34] Kanjanapothi D, Tejasen P. Chiang Mai Med Bull 1971; 10:89-97.
- [35] Hattori M, Nakabayashi T, Lim YA, Miyashiro H, Kurokawa M, Shiraki K, Gupta MP, Correa M, Pilapitiya U. Phytother Res 1995; 9(4):270-276.
- [36] Deshmukh SK, Jain PC., Agrawal SC. Fitoterapia 1986; 58(4):295-297.
- [37] Banerji R, Srivastava AK, Misra G, Nigam SK, Singh S, Nigam SC, Saxena RC. Indian Drugs 1979; 17:6-8.
- [38] Sato Y, Oketani H, Singyouchi K, Ohtsubo T, Kihara M, Shibata H, Higuti T. Biol Pharm Bull 1997; 20(4):401-404.
- [39] Chung TH, Kim JC, Lee CY, Moon MK, Chae SC, Lee IS, Kim SH, Hahn KS, Lee IP. Phytother Res 1997; 11(3):179-182.
- [40] Cheng HY, Lin TC, Yu KH, Yang CM, Lin CC. Biol Pharm Bull 2003; 26(9):1331-1335.
- [41] Lee HS, Won NH, Kim KH, Lee H, Jun W, Lee KW. Biol Pharm Bull 2005; 28(9):1639-1644.
- [42] Kumazawa S, Taniguchi M, Suzuki Y, Shimura M, Kwon M, Nakayama T. J Agric Food Chem 2002; 50(2):373-377.
- [43] Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Free Radic Biol Med 1999; 26:1231-1237.
- [44] Bajpai M, Pande A, Tewari SK, Prakash D. Int J Food Sci Nutr 2005; 56(4):287-291.
- [45] Shui G, Leong LP. J Chromat A 2004; 1022(1): 67-75.