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Effect of *Citrus hystrix* and *Citrus limon* extracts on antibacterial activity against human pathogens.

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ABSTRACT

Objective: To isolate bacteria from infected human and also screening of antibacterial property against human pathogenic bacteria in two species of Citrus. Methods: The present study was isolated microorganism from human pathogens and determined by Agar diffusion method. Results: The results showed that the aqueous extract of bark, leaf and fruit peels of the two species did not produce any significant result on tested viz, Staphylococcus aureus, Salmonella typhi, Escherichia coli, Klebsiellla pneumonia and Proteus vulgaris microorganisms. However the methanolic extracts of Citrus hystrix fruit peel produced the maximum inhibition zone of 22 mm and 19 mm against Staphylococcus aureus and Salmonella typhi respectively. Citrus hystrix fruit peel extracts has produced the inhibition zone of 15 mm and 11 mm against Escherichia coli and Klebsiellla pneumonia respectively. The fruit peel of Citrus limon produced an maximum inhibition zone of 20 mm and 18 mm against Staphylococcus aureus and Salmonella typhi respectively. The methanol extract of *Citrus limon* produced an maximum inhibition zone of 13 mm and 9 mm against Escherichia coli and Klebsiellla pneumonia respectively. But Proteus vulgaris was minimum level of inhibition by all the solvent extract viz., bark, leaf and fruit peels of two species of Citrus. Conclusions: The present investigation conclude that the extract methanolic fruit peel extract of Citrus hystrix and Citrus limon possess antibacterial properties that can be explored as a viable, alternative source to commercially available antibiotic drugs.

1. Introduction

Nature has been a source of medicinal agents and an impressive number of modern drugs. Higher plants have been a rich source of medicine because they produce a wide array of bioactive molecules. Each plant and plant parts has one or more substances that can be used for therapeutic purposes and the plants are called medicinal plants. Medicinal plant substances can inhibit growth of pathogens or kill them because they have antimicrobial compound. The basis of sophisticated traditional medicine system and natural products provide excellent leads for new drug development (Newman *et al.*, 2000).

In recent years drug resistance to human pathogenic

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bacteria has been commonly reported from all over the world. Microorganisms are present on almost all the body surface both externally and internally. But their disease causing potential depends upon the virulence, *i.e* ability to cause a disease. The indigenous microflora of human body contains pathogens mainly Staphylococcus members. It causes urinary tract infections and many other diseases. During the last 10 years the development of new antimicrobial drugs have been established because the society has been facing one of the most serious public health care over the emergence of infectious bacterial displaying resistance to many drugs. There is a continuous and urgent need to discover the new antimicrobial compounds with diverse chemical structures and novel mechanism of an action because there has been an alarming increase in the incidence of new and re-emerging infectious diseases. Another big concern is the development of resistance to the antibiotics.

Citrus is one of the most important commercial fruit

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crops grown in all continents of the world [Tao et al., (2008)]. Importance of Citrus is attributed to its diversified use and growing world demand with about 102.64 million tones total world production and probably stands first largest among the produced fruit [NAQVI (2004)]. The Citrus peels are rich in nutrients and contain many phytochemicals, they can be efficiently used as drugs or as food supplements too. Since there is an increase in the number of antibiotic resistance pathogens, there is always a search of an alternative drug that is regarded as safe. Citrus peels if proved to have antibacterial activity; which generates large peel wastes that can also be used in food industry as preservative. It is well known that essential oils from *Citrus* spp. have pronounced antimicrobial effect against both bacteria and fungi (Lanciotti et al., 2004). Citrus hystrix peel oil contained terpinen-4 (13.0%), pinene (10.9%), terpineol (7.6%), 1,8-cineole (6.4%), citronellol (6.0%) and limonene (4.7%) (Waikedre et al.,2010.) It is already reported that ethyl acetate extract of Citrus hystrix peel had stronger antibacterial activity (Chanthaphon et al., 2008). The antioxidant activities of citrus species are in accordance with their amount of phenolics. C. hystrix contained high phenolic content compared to other Citrus species, which was responsible for its high antioxidant activity. Several reports showed a close relationship between total phenolic content and high antioxidant activity (Prasad et al., 2005; Amin et al., 2006; Li et al., 2009).

2. Materials and methods

2.1 Plant material

The bark, leaves and fruit peel of *Citrus limon* (L.) Burm.f. and *Citrus hystrix* DC. were collected from Mayiladuthurai, $(11 \circ 6' 35'' N, 79 \circ 39' 0'' E)$ Nagapattinam District TamilNadu, India, during April, 2010. Two species of *Citrus* were identified and conformed in Rabinat herbarium, St. Joseph's College, Trichirapalli. India. The collected specimens were identified with the help of Flora of Presidency of Madras (Gamble, 1953). Species identification was confirmed by comparing the collected specimens with the Herbarium of Department of Botany, University of Calicut, Kerala, India. Duplicated voucher specimens have been deposited in the herbarium in Department of Botany, Annamalai University, Tamil Nadu, India.

2.2.EXTRACT PREPARATION

The bark, fresh leaves and fruit peels were shade dried separately and ground into fine powder $(20_{mesh}; \sim 1 \text{ g})$ with mechanical grinder Thirty grams of bark, leaf and fruit peel powder was then macerated in 100 ml absolute Methanol for 48 h by soxhlet extraction apparatus. The extract was evaporated to dryness at 40 $^{\circ}$ C in a vacuum using a rotary evaporator and store at 5 $^{\circ}$ C in a refrigerator. The aqueous

extract, the bark, leaves and fruit peel were shade dried and ground to fine particles with a mechanical grinder. It was then macerated in 500 ml of sterile distilled water for 48 h using a 500 ml conical flask. The conical flask was properly labelled and covered with aluminum foil to prevent contamination. The extracts were then filtered off with sterile filter paper (Whitman No 1). The prepared extract was evaporated to dryness and stored in the refrigerator at 5 \degree C for further use.

2.3. Isolation of microorganisms

The test microorganisms like *Staphylococcus aureus*, *Salmonella typhi, Escherichia coli, Klebsiellla pneumonia*e and *Proteus vulgaris* were isolated from infected human skin, mouth and urine at Government Hospital, Trichirapalli. The samples were plated out on Blood Agar (Himedia) for prepared according to manufacturer's specifications. The isolated microorganism was subcultured and characterized using the methods of Cruickshank *et al.* (1975) and Cowan (1985). They were then stored in agar slants in the refrigerator at 4 \degree C.

2.4. Antimicrobial susceptibility test

The isolated bacterial cultures were maintained on nutrient agar slants at 40C. These bacterial cultures were diluted using Nutrient Broth and diluted bacterial culture (0.2ml) was spread over sterile Nutrient agar plates. About 0.2ml of the plant extracts were applied for the sterile filter paper disc (Whatman No.1, 6mm in diameter) before being placed to the agar plates. Each extracts was tested triplicate. The plates were incubated at 370C for 24 hours. The inhibition zones were recorded. The antimicrobial activities of plant extracts were indicated by clear zones of growth inhibition. The commercial Ampicillin antibiotic disc (1 mg/mL-1, Himedia) was used as control disc.

3. Results

Table 1 showed that the results of antibacterial activity of extract from bark, leaves and fruit peel of *Citrus hystrix* and *Citrus limon*.

Table 1

Antibacterial activity on methanol extracts of *Citrus hystrix* and *Citrus limon* by disc Diffusion method

Plant	Parts	Inhibition zone (mm)*				
		S. aureus	K. pneumonia	S. typhi	E. coli	P. vulgaris
С.	Bark	14±0.3	6.0±0.2	10±0.2	8.5±0.2	3.0±0.1
hystrix	Leaf	18±0.5	7.0 ± 0.2	13±0.3	11±0.3	5.0 ± 0.1
	Fruit peel	22±0.5	11±0.3	18 ± 0.5	15 ± 0.5	8.0±0.2
С.	Bark	12±0.3	5.0 ± 0.2	08±0.2	07±0.2	2.0±0.1
limon	Leaf	15±0.4	7.5 ± 0.2	11±0.3	09±0.3	4.0 ± 0.1
	Fruit peel	19±0.5	9.0±0.3	16±0.5	13±0.5	6.0±0.2

mm* = Mean of three replicates

The commercial Ampicillin antibiotic disc (1 mg/mL-1, Himedia) was used as positive control. The extractions were carried out using methanol and aqueous solvents. The methanol extracts gave the high activity against all tested bacteria but the aqueous extract has not any antibacterial activity. Methanol extract inhibited the growth of five tested bacteria and the maximum inhibition zone was recorded against *Staphylococcus aureus* and *Salmonella typhi*. The methanol extract of C. hystrix fruit peel extract was highest inhibition zone (Tab. 1 & Fig. 1) compared to *Citrus limon* fruit peel extract. However the methanolic extracts have shown (Tab 1 & Fig. 1) fruit peel of *Citrus hystrix* produced an inhibition zone of 19 mm and 22 mm against *Salmonella typhi* and *Staphylococcus aureus* respectively.

Antibacterial activity on methanol extract of Citrus hystrix



Figure 1. Antibacterial activity on methanol extract of *Citrus hystrix* by Disc diffusion Method

Extract from fruit peel of *Citrus hystrix* produced highest inhibition zone compared to control against *Staphylococcus aureus*. Whereas the fruit peel extract of *Citrus hystrix* (Tab. 1 & Fig. 2) produced an inhibition zone of 14 mm and 16 mm on Klbesiella pneumonia and Escherichia coli respectively. Citrus peel extracts showed a significant antibacterial activity against all the test organisms compared to other like leaf and bark extracts. The methanol solvent shows that *Citrus hystrix* and *Citrus limon* fruit peel extract has high degree of antibacterial activity as compared to the leaf and bark extract.

Antibacterial activity on methanol extract of Citrus hystrix



Figure 2 Antibacterial activity on methanol extract of *Citrus limon* by Disc diffusion Method

Table 2

Positive control use against tested bacteria

Micro organism	Control Inh. zone (mm)
Staphylococcus aureus	20
Klbesiella pneumonia	25
Salmonella typhi	35
Escherichia coli	35
Proteus vulgaris	20

Control = Ampicillin (1 mg/mL-1)

4. Discussion

The methanol extract may have diverse antibacterial agent that has different modes of action or the bacteria may have a special metabolism to overcome or adapt its activity. So, methyl alcohol proves to be a good solvent for the extraction of antibacterial agents from both sources as it has shown better yield as well as antibacterial activity. Aqueous extract showed no any degree of sensitivity against tested pathogenic bacteria and was not able to inhibit the growth of all of the tested pathogens. Ashokumar et al reported that, the difference may be in the phytochemical composition in various part of the plant or may be also due to the extraction method used and or environmental factors or difference in the genotypes of the Citrus plant used [5]. The citrus peel extract exhibited similar or higher antibacterial activity.

The present experimental study concludes that, the methanolic fruit peel extract of *Citrus hystrix* and *Citrus limon* possess antibacterial effect on pathogenic bacteria such as *Staphylococcus aureus*, *Salmonella typhi*, *Escherichia coli*, Klbesiella pneu*Proteus vulgaris*. The results of the present study suggest that the extract methanolic fruit peel extract of *Citrus hystrix* and *Citrus limon* possess antibacterial properties that can be explored as a viable, alternative source to commercially available antibiotic drugs. Further studies are needed to test it on other microorganisms and against various infections, where in the information procured would further serve as a strong evidence for the plant as potent antimicrobial agent.

Conflict of interest statement

We declare that we have no conflict of interest.

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References

- Srisukha V, Tribuddharatb C, Nukoolkarne V, Bunyapraphatsarae N, Chokephaibulkitd K, Phoomniyomb S, Chuanphungb S, Srifuengfung S. Antibacterial activity of essential oils from Citrus hystrix (makrut lime) against respiratory tract pathogens. *ScienceAsia* 2012; 38: 212–217
- [2] Verma V, Singh R, Tiwari R K, Srivastava N and Verma A. Antibacterial activity of extracts of Citrus, Allium & Punica against food borne spoilage. Asian Journal of Plant Science and

Research, 2012; 2 (4):503-509

- [3] Upadhyay RK, Dwivedi P, Ahmad S. Screening of antibacterial activity of six plant essential oils against pathogenic bacterial strains. *Asian J. Med. Sci.*, 2010; 2: 152–158.
- [4] Dhanavade J M, Jalkute B C, Ghosh C J and Sonawane D. Study Antimicrobial Activity of Lemon (Citrus lemon L.) Peel Extract. British Journal of Pharmacology and Toxicology 2011; 2(3): 119–122.
- [5] Ashok kumar K., Narayani, M., Subanthini A and Jayakumar M.. Antimicrobial Activity and Phytochemical Analysis of Citrus Fruit Peels –Utilization of Fruit Waste. *International Journal of Engineering Science and Technology* 2011; 3(6): 5414–5421.
- [6] Waikedrea J, Dugayb A, Barrachinac I, Herrenknechte C, Cabaliona P, and Fournet A. Chemical Composition and Antimicrobial Activity of the Essential Oils from New Caledonian *Citrus macroptera* and *Citrus hystrix*. *Chemistry & Biodiversity* 2010; 7: 871–877
- [7] Uchechi N. Ekwenye and Oghenerobo V. Edeha.. The antibacterial activity of crude leaf extract of *Citrus sinensis* (sweet orange), *International Journal of Pharma and Bio Sciences*, 2010;1(4):742-750.
- [8] Cruickshank R, Duguid JP, Marmion BP, Swain RH (1975). Medical Microbiology (12th ed) Longman group Ltd. Edinburgh, London pp. 180–188
- [9] Doughari, J. H, El-mahmood, A. M. and Tyoyina, I (2008). Antimicrobial activity of leaf extracts of Senna obtusifolia (L). African Journal of Pharmacy and Pharmacology 2(1): 007-013
- [10] Ekpo M. A. and Etim P. C.(2009). Antimicrobial activity of ethanolic and aqueous extracts of Sida acuta on microorganisms from skin infections. Journal of Medicinal Plants Research 3(9): 621–624
- [11] Elizabeth K.M (2005). Antimicrobial Activity Of Terminalia Bellerica Indian Journal of Clinical Biochemistry 20 (2): 150–153
- [12] Doreen S.H., Laili C. Rose1, Suhaimi1 H, Mohamad1 H, Mohd Z. H.,Rozaini1 and Mariam Taib. Preliminary evaluation on the antibacterial activities of citrus hystrix oil emulsions stabilized by tween 80 and span 80. Int J Pharm Pharm Sci, 2011;3(2):209–211
- [13] Leite S.P, Vierira J.R.C, Medeiros P.L, Leite R.M.P, Lima V.L, Xavier H.S and Lima E.O (2006). Antimicrobial Activity of Indigofera suffruticosa. CAM :1-5.
- [14] Najjaa .H, Ammar .E and Neffati .M (2009). Antimicrobial activities of protenic extracts of Allium roseum L., a wild edible species in North Africa. Journal of Food, Agriculture & Environment (7): 1 5 0 - 1 5 4.
- [15] Javed S, Javaid A, Mahmood Z, Javaid A and Nasim F. Biocidal activity of citrus peel essential oils against some food spoilage bacteria. *Journal of Medicinal Plants Research*, 2011; 5(16): 3697–3701
- [16] Rahman and Mohammad Junaid (2008). Antimicrobial Activity Of Leaf Extracts Of Eupatorium Triplinerve Vehl. Against Some Human Pathogenic Bacteria And Phytopathogenic Fungi. Bangladesh J. Bot. **37**(1): 89–92
- [17] Rahman M, M. Mominul Islam Sheikh, Shamima Akhtar Sharmin,
 M. Soriful Islam, M. Atikur Rahman1, M. Mizanur Rahman and M.
 F. Alam1 (2009). Antibacterial Activity of Leaf Juice and Extracts

of Moringa oleifera Lam. against Some Human Pathogenic Bacteria.. CMU. J. Nat. Sci. **8**(2): 219

[18] Wei L.S, Najiah Musa, Chuah Tse Sengm, Wendy Wee and Noor Azhar Mohd Shazili (2008). Antimicrobial properties of tropical plants against 12 pathogenic bacteria isolated from aquatic organisms. African Journal of Biotechnology 7 (13): 2275-2278