

Antimicrobial Activity of the Masticatory *Cola acuminata* Nut (Gooro)

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Abstract: The aim of this study was to investigate the possible inhibitory activity of *Cola acuminata* seed (Petroleum ether, methanolic and aqueous) extracts against two standard Gram positive (*Bacillus subtilis* and *Staphylococcus aureus*), three Gram negative (*Escherichia coli*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*) bacteria and one fungus (*Candida albicans*). Petroleum ether, methanolic and aqueous extracts of *Cola acuminata* nuts at concentrations of 10, 12.5, 25 and 50% were screened for antimicrobial activity against two standard Gram positive (*Bacillus subtilis* and *Staphylococcus aureus*), three Gram negative (*E. coli*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*) bacteria and one fungus (*Candida albicans*). No inhibitory activity was detected using all concentrations of petroleum ether extract for all tested microorganisms. Wide range of activity was exhibited by both aqueous and methanol extract. However, the former highest activity was detected against *Staphylococcus aureus* at the two concentration 25 and 50% (20 and 24 mm) while the latter was found against *Candida albicans* at the concentration of 25% (20 mm).

Key words: Antimicrobial activity, *Bacillus subtilis*, *Candida albicans*, *Cola acuminata*, *E. coli*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*

INTRODUCTION

The demand for safer and natural foods has been increasing since consumers have become more concerned over the presence of chemical residues in their stuff. There is however, a strong demand for the reduction or elimination of the synthetic chemical. It has long been recognized that some plant materials exhibit antimicrobial properties. The use of these plant materials as preservatives and as means of preventing microorganism development in foods has become the subject of extensive studies (Gould, 1996).

In particular, the inhibitory effects of extracts of many kinds of herbs and spices against food borne bacteria and pathogens have been reported. Among these are cassia, clove, garlic, sage and thyme (Shelef, 1983; Zaika and Kissinger, 1981; Saleem and Ai-Delaimy, 1982; Tassou *et al.*, 2000).

Currently there is growing demand worldwide of consumers to minimize chemical preservation that can be detrimental to human health (Cho *et al.*, 1995; Smid and Gorris, 1999). Consequently, spices, herbs and naturally occurring phenolics from various plant sources are being studied in detail in response to consumer requirements for fresher and more natural additive-free products (Nychas, 1995; Tassou *et al.*, 1997).

Cola, a member of the chocolate family Sterculiaceae (from which nuts are obtained) a tree probably originally from western tropical Africa (Encyclopedia Americana, 1988). The cola nut is a capsule-shaped fruit composed of fleshy, irregularly shaped seeds. The seeds of *Cola acuminata* are pink, red or white when fresh, and become brown and hard once they are dried. Their bitter and astringent taste is why these seeds are called nuts. Chewing of the fresh *Cola acuminata* nut is a wide spread habit of the Western and sub-Saharan indigenous populations of Africa especially in Northern Nigeria, Ivory Coast, Sierralione Liberia and Sudan (Russell, 1955; Purselove, 1968; Rosengarten, 1984), while the dried nuts are used beverages and pharmaceutical purposes in Europe North America (Ayodele, 1995).

The *Cola acuminata* nut's primary social and economic significance lies in its being a very concentrated source of Central Nervous System (CNS) stimulant, caffeine which may help in relieving migraine (Kiple and Ornelas 2000b), Theobromine which act as cerebral vasodilator and thought to relieve pain and neuralgia (Hirt and M'pia, 2001) and proanthocyanidin is used as anti-trypanosoma compound effective against *Trypanosoma brucei* (Kubata *et al.*, 2005). They also ease hunger and thirst, eliminate fatigue, provide energy by stimulating the muscles and nerves and enhance intellectual activity

Table 1: Differential characteristics of test bacteria

Characteristics	<i>S. aureus</i>	<i>E. coli</i>	<i>Ps. aeruginosa</i>	<i>B. subtilis</i>	<i>Klebsiella</i> spp
Gram stain	+ve	-ve	-ve	+ve	-ve
Motility	+ve	+ve	+ve	+ve	-ve
Catalase	+ve	+ve	+ve	+ve	+ve
Nitrate	+ve	+ve	+ve	+ve	+ve
Arginine	+ve	+ve	+ve	+ve	-ve
Indole	-ve	+ve	-ve	-ve	-ve
Methyl red	-ve	+ve	+ve	-ve	-ve
Voges-Proskauer	+ve	-ve	-ve	+ve	+ve
Gas production from glucose	-ve	+ve	+ve	-ve	+ve
Acid production from glucose	+ve	+ve	-ve	-ve	+ve
Acid production sucrose	-ve	+ve	+ve	+ve	-ve
Acid production from lactose	+ve	+ve	-ve	+ve	+ve
Oxidase	-ve	+ve	+ve	+ve	-ve
Urease	+ve	-ve	+ve	+ve	+ve

+ve = 90% or more strains had positive reaction; -ve = 90% or more strain had negative reaction; -: Non- determined

(Sundstrom, 1966; Nickalls, 1986). Traditionally, the leaves, flowers, fruit follicles and park of *Cola acuminata* were used to prepare a tonic as a remedy for dysentery, cough, diarrhea, and vomiting (Ayensu, 1978).

MATERIALS AND METHODS

Plant materials: *Cola acuminata* seeds were purchased from a local market in Khartoum, authenticated by the scientists at the Aromatic and Medicinal Plants Research Institute, Khartoum, and brought to the research laboratory, University of El Neelain, Khartoum, Sudan. The plant tissues were cleaned, shade-dried, powdered by a mechanical grinder.

Standard microorganisms: The test organism used in this study were kindly provided by the scientists at the National Health Institute, Khartoum and designated as follows:

Bacteria:

Bacillus subtilis- NCTC/8236
Staphylococcus aureus- ATCC/25923
Escherichia coli- ATCC/27853
Klebsiella spp- ATCC/3565
Pseudomonas aeruginosa- ATCC/2785
 NCTC National Collection Type Culture, London, UK.
 ATCC American Type Culture Collection, Manassas, AV, USA.

Fungus:

Candida albicans

Antibiotics:

Gentamicin (Roussel Laboratories Ltd, England)
 Nystatin (Sigma Chemical Company, USA)

Cultural media: Standard media, nutrient broth (Oxoid Ltd., London) form the base of most of the media used. Nutrient agar (Oxoid Ltd., London) was used to prepare the enriched culture media.

Mueller- Hinton agar (Oxoid, Ltd, London), used for all antibiotic sensitivity test as standard drug as well as for plant extracts evaluation.

Sabouraud's Dextrose agar (Difco, USA), used as enriched culture media for fungi

Preparation of crude extracts: Twenty grams of powdered *Cola acuminata* seeds were extracted successively with petroleum ether 90% at 37°C and methanol 95% at 37°C using Soxhlet apparatus extractor for 3 h. The extracts were evaporated under reduced pressure, air-dried and yields were recorded. The aqueous extract was dried by freeze dryer and weighed. The extracts of the seeds were reconstituted at the time of testing in concentrations of 50, 25, 12.5 and 10%.

Preparation of stock extracts solution: One grams of each extract was dissolved in 1 ml. of the same solvent used for extraction.

Preparation of the test organism: The properties at the standard bacteria are summarized in (Table 1).

Staphylococcus aureus: Gram-positive, non-motile, non-spore forming, aerobic, facultatively anaerobic organism, 0.5 mm in diameter. It occurs singly, in pairs or irregular and many of the strain form a golden yellow pigment on colony of good growth on ordinary media. Catalase and urease positive, oxidase and indol negative.

Escherichia coli: Gram-negative rod, 1.1-1.5 mm wide and 2.0-6.0 mm long with rounded ends and shape varying from coccoid to rod, motile, aerobic, facultatively anaerobic oxidase and urease negative, citrate can not be used as a sole carbon, most strain are fermenters of methyl red positive, catalase and indol positive and VP negative.

Pseudomonas aeruginosa: Gram-negative bacillus, non-spore forming, non- capsulated, motile by one or two polar flagella aerobic, facultatively anaerobic, grow on a wide variety of culture media, catalase positive.

Table 2: Evaluation of antimicrobial activity of *Cola acuminata* seed petroleum ether, methanol and aqueous extracts

Organism	Extract conc. %	Inhibition zone (mm)		
		Petroleum ether extract	Methanol extract	Aqueous extract
<i>Bacillus subtilis</i>	10	(-)	14-15	11-12
	12.5	(-)	10-12	14
	25	(-)	18	14
	50	(-)	16	18
	10	(-)	12-12.5	12
<i>Staphylococcus aureus</i>	12.5	(-)	13	10
	25	(-)	15	22
	50	(-)	17	24
	10	(-)	11-12	10-14
<i>Escherichia coli</i>	12.5	(-)	9	8
	25	(-)	10	9
	50	(-)	11	11
	10	(-)	14-15	14-15
<i>Klebsiella pneumoniae</i>	12.5	(-)	11	10
	25	(-)	12-13	9
	50	(-)	9-10	10
	10	(-)	12-14	11-12
<i>Pseudomonas aeruginosa</i>	12.5	(-)	11	10
	25	(-)	11-12	11
	50	(-)	18	12
	10	(-)	13-14	13-15
<i>Candida albicans</i>	12.5	(-)	14	14
	25	(-)	20	9
	50	(-)	18	8

(-) = No inhibition zone was observed

Bacillus subtilis: *Bacillus subtilis*, known also as the hay bacillus or grass bacillus, is a Gram-positive, catalase-positive bacterium commonly found in soil. A member of the genus *Bacillus*, *B. subtilis* is rod-shaped, and has the ability to form a tough, protective endospore, allowing the organism to tolerate extreme environmental conditions. Unlike several other well-known species, *B. subtilis* has historically been classified as an obligate aerobe, though recent research has demonstrated that this is not strictly correct.

Klebsiella pneumoniae: Gram-negative, non-motile straight rods, 0.3-1.0×0.06-6.0 µm, arranged singly, in pairs or in short chain; often surrounded by a capsule, facultatively anaerobic, having both a respiratory and a fermentative metabolism.

Antimicrobial assay: The antimicrobial activity of the extract was assessed by the agar-well diffusion method (Kingsbury and Wagner, 1999). The nutrient agar medium was properly and separately inoculated with the standard organism at 10⁶ cfu/mL to achieve confluent growth and allow drying at room temperature. On each inoculated plate, 10 mm-diameter wells were bored in the agar using a sterile cork borer. Hundred µl aliquot of each concentration (50, 25, 12.5 and 10%) was reconstituted extract and placed into the well using a standard Pasteur pipette. The agar was allowed to solidify before incubation at 37°C for 18 h for bacteria and 25°C for 72 h for the yeast.

The diameter of clear zones around each well was measured (mm) after incubation to determine the strength

of inhibition. Three replicates were made from each concentration and comparative activity was recorded. The antimicrobial activity of the plant extract against the standard microorganism was evaluated and compared with that of Gentamicin (Bacteria) and Nystatin (Yeast).

RESULTS AND DISCUSSION

Extract yield: The 50 g of *Cola acuminata* powdered seed yielded 0.9, 4.8 and 6 g when extracted by petroleum ether, methanol and water respectively.

Evaluation of in vitro antimicrobial activity of petroleum ether, methanol and aqueous extracts of Cola acuminata seeds: The inhibitory activity of the different *Cola acuminata* extracts are presented in Table 2. Results indicated that the petroleum ether extract had no inhibitory activity against both bacteria and yeast. The methanolic and aqueous extracts of *Cola acuminata* seeds exhibited a wide range and a broad-spectrum antimicrobial activity against all the test bacteria and fungi ranging from 8-20 and 8-24 mm, respectively and this was compared well with Gentamicin and Nystatin (Table 3, Fig. 2). The highest inhibitory effect of the aqueous extract was observed against *Staphylococcus aureus* at the concentrations of 25 and 50% (Fig. 1), whereas the 25% concentration of the methanolic extract showed the highest inhibitory effect against *Candida albicans* (20 mm). The methanolic extract of *Kigelia africana* exhibited high activity than aqueous and chloroform extracts against *Salmonella typhi* but moderate activity against *Staphylococcus aureus*. Although, it was



Fig.1: Inhibition zone produced by methanol extract (10, 12.5, 25, 50%) of *Cola acuminata* seeds against *Staph. aureus*

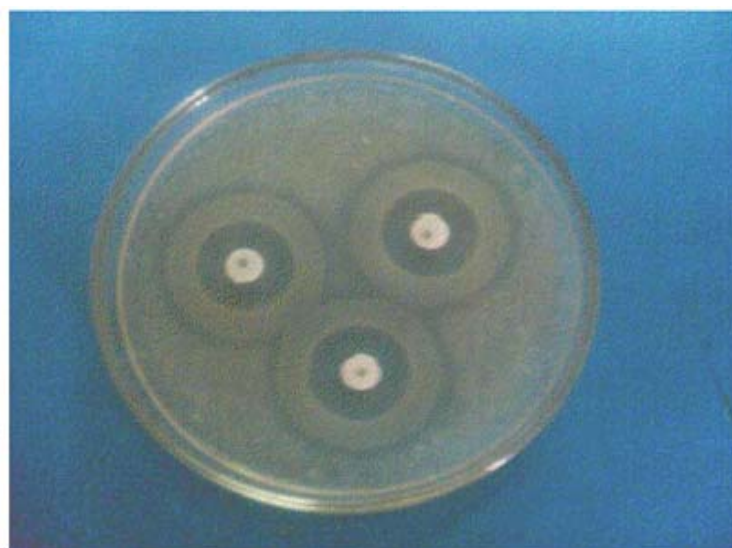


Fig. 2: Inhibition zone produced by Gentamicin against *Staph aureus*

Gentamicin (10 mg/mL)	Inhibition zone (mm)
<i>B. subtilis</i>	18
<i>S. aureus</i>	24
<i>E. coli</i>	20
<i>Kl. Pneumoniae</i>	17
<i>Ps. aeruginosa</i>	15
Nystatin (10 mg/mL)	
<i>Candida albicans</i>	14

reported that methanol was a better solvent for the consistent extraction of antimicrobial substances from medicinal plants when compared to other solvents such as aqueous, ethanol, chloroform and hexane (Lin *et al.*,

1999; Ahmed *et al.*, 1998; Eloff, 1998), the methanolic extract of *Parinariium glaberrimum* Hassk fruit, seed and peel did not show any antimicrobial activity when examined against a wide range of bacteria and fungi including *Staphylococcus aureus*, *E. coli*, *Bacillus subtilis* and *Candida albicans* (Moniharapon and Hashinaga, 2004). The methanol extract of *Kigelia africana* (belonging to the family Bignoniaceae) presented a higher activity than the aqueous and chloroform extracts with moderate activity against *Staph aureus* and *E. coli*.

The antimicrobial activity may involve complex mechanisms like the inhibition of the synthesis of cell

wall, cell membrane, nucleic acids and protein as well as the inhibition of the metabolism of nucleic acids (Oyaizu *et al.*, 2003).

The antibacterial activity of plant extracts can be attributed not only to a single bioactive principle but also in concert action with other compounds (Sunayana *et al.*, 2003). A number of phytochemicals have been studied for their antimicrobial activity and found potentially useful against infectious diseases. The chemical structure of the antimicrobial agents found in higher plants belong to most commonly encountered classes of higher plants secondary metabolites such as flavonoids (Watchter *et al.*, 1999), terpens (Conveney *et al.*, 1985), terpenoids (Osawa *et al.*, 1990; Habibi *et al.*, 2000) and phenolic acids (Fernandez *et al.*, 1996).

CONCLUSION

From this study it was concluded that the spectrum of antimicrobial activity of the investigated methanolic and aqueous extracts of *Cola acuminata* seed against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* in addition to the yeast *Candida albicans*, was found comparable to the reference antibacterial (Gentamicin) and antifungal (Nystatin) drugs. The petroleum ether extract of the same plant was found to have no inhibitory activity against both tested bacteria and yeast.

This investigation may justify the use of this plant in the local traditional medicine. Detailed *in vitro* studies utilized the characterized active principle of the plant are necessary for assessing these antimicrobial constituents.

ACKNOWLEDGMENT

The authors would like to thank Mr. El-Khatim Hassan Abdelgadir and all the teaching staff of the School of Biotechnology, Faculty of Science and Technology, University of El Neelain for their valuable technical assistance.

REFERENCES

Ahmed, I., Z. Mohmood and F. Mohammad, 1998. Screening of some Indian medicinal plants for their antimicrobial properties. *J. Ethnopharmacol.*, 62: 183-193.

Ayensu, E.S., 1978. Medicinal plants of West Africa. Reference Publication International, Michigan, Michigan.

Ayodele, E.A., 1995. Physico-chemical properties of some kola (*Cola nitida*) growing soils of Nigeria. *Nigerian J. Tree Crop Res.*, 1(2): 37-51.

Cho, S.H., S.Y. Lee, J.W. Kim, G.H. Ko and I.W. Seo, 1995. Development and application of natural antimicrobial agents isolated from grape-fruit seeds extract, antimicrobial activities of grape fruit extract. *J. Food Hyg. Saf.*, 10: 33-39.

Conveney, D., N. Fakuda, J. Reilly, J. Polonsky, T. Prange, D. M. X. Donnelly and F. Aba, 1985. Antibacterial sesquiterpenes aryl esters from *Armillaria mellea* *J. Natural Prod.*, 48:10-11.

Eloff, J.N., 1998. Which extracts should be used for the screening and isolation of antimicrobial components from plants. *J. Ethnopharmacol.*, 60: 1-8.

Encyclopedia Americana, 1988. Encyclopedia Americana, CT: Grolier, 7: 216.

Fernandez, M.A., M.D. Garcia and M.T. Saenz, 1996. Antibacterial activity of the phenolic acid fractions of *Scrophularia frutescens* and *Scrophularia sambucifolia*. *J. Ethnopharmacol.*, 53: 11-14.

Gould, G.W., 1996. Industry perspectives on the use of natural antimicrobial and inhibitors for food application. *J. Food Prot.*, 59: 82-86.

Habibi, Z., F. Eftekhari, K. Samiee and A. Rustaiyan, 2000. Structure and antibacterial activity of 6 new labdane diterpenoid from *Salvia leriifolia*. *J. Natural Prod.*, 63: 270-271.

Hirt, H.M. and B. M'pia, 2001. Natural Medicine in the Tropics. Anamed. 2nd Edn., Drucherei Bauer, Winnenden.

Kingsbury, D.T. and G.E. Wagner, 1999. Microbiology. Williams and Wilkins, Maryland.

Kiple, K.E. and K.C. Ornelas, 2000b. Kola nuts in: The Cambridge World History of Food. Cambridge University Press, New York, NY, pp: 684-692.

Kubata, B.K., K. Nagamune, N. Muramkami, P. Merkel, Z. Kabututu, K. Samuel, Martin, M. Taba, Kalulu, H. Mustakuk, M. Yoshida, M. Ohnishi, K.T. Kameyama, M. Duzenko and Y. Urade, 2005. *Cola acuminata* proanthocyanidins: a class of anti-trypanosomal compounds effective against *Trypanosoma brucei*. *Int. J. Parasitol.*, 35: 91-103.

Lin, J., W.A.R. Opak and M. Geheeb-Keller, 1999. Preliminary screening of some traditional medicinal plants for anti-inflammatory and antimicrobial activities. *J. Ethnopharmacol.*, 68: 267-274.

Moniharapon, E. and F. Hashinaga, 2004. Antimicrobial activity of Atung (*Parinarium glaberrimum* Hassk) fruit extract. *Pak. J. Biol. Sci.*, 7(6): 1057-1061.

Nickalls, R.W.D., 1986. W.F. Daniell (1917-1965), and the discovery that the cola-nuts contain caffeine. *The Pharm. J.*, 236: 401-402.

Nychas, G.J.E., 1995. Natural Antimicrobial from Plants. In: Gould, G.W., (Ed.), New Methods of Food Preservation Blackie Academic and Professional, London, pp: 58-89.

Osawa, K., T. Matsumoto, T. Marnyama, T. Takiguchi, K. Okuda and I. Takazoe, 1990. Studies of antimicrobial activity of plant extracts and their constituents against periodontopathic bacteria. *Bull. Tokyo, Dental Collage*, 31: 17-21.

Oyaizu, M., Y. Fujimoto, H. Ogihara, K. Sekimoto, A. Naruse and U. Naruse, 2003. Antioxidative and antimicrobial activities of extracts from several utility plants. *Food Preserv. Sci.*, 29: 33-38.

- Purseglove, J.W., 1968. Tropical Crops: Dicotyledons. Longmans Green and Co. Ltd., London.
- Rosengarten, F., 1984. The Book of Edible Nuts. Walker and Company, New York.
- Rusell, T.A., 1955. The Cola of Nigeria and Cameroon. *Trop. Agric.*, 32: 210-240.
- Saleem, Z.M. and K.S. Ai-Delaimy, 1982. Inhibition of *Bacillus cereus* by garlic extract. *J. Food Prot.*, 45: 1007-1009.
- Shelef, L.A., 1983. Antimicrobial effect of spices. *J. Food Prot.*, 6: 29-44.
- Smid, E.J. and G.M. Gorris, 1999. Natural Antimicrobials for food Preservation. In: Rahman, M.S. (Ed.), Handbook of Food Preservation, Marcel Dekker. New York, pp: 285-308.
- Sunayana, V., A. Vadivukkarasi, T. Rajendran, X. Francis and E. Natarajan, 2003. Antibacterial potential of *Plectranthus amboinicus* (Lour) Spreng. A study *in vitro*. *J. Swamy Botanic. Club*, 20: 55-58.
- Sundstrom, L., 1966. The Cola nuts. Function in West African Social Life. *Studia Ethnographia Upsaliensia*, 26: 135-145.
- Tassou, C.C. and E.H. Drosinos, 1997. Wak antimicrobial effect of carob (*Ceratonia siliqua*) extract against food-related bacteria in culture media and model food system. *World J. Microbiol. Biotechnol.*, 13: 479-481.
- Tassou, C., K. Koutsoumanis and G.J.E. Nychas, 2000. Inhibition of *Salmonella enteritidis* and *Staphylococcus aureus* in nutrient broth by mint essential oils. *Food Res. Intl.*, 33: 273-280.
- Watchter, G., A. Hoffmann, T. Furbacher, M.E. Blake and B.N. Timmerman, 1999. Antibacterial and antifungal flavanones from *Eysenhardtia texana*. *Phytochem.*, 52: 1469-1471.
- Zaika, L.L. and J.C. Kessinger, 1981. Inhibitory and stimulatory effects of oregano on *Lactobacillus plantarum* and *Pediococcus cerevisiae*. *J. Food Sci.*, 46: 1205-1210.