A Review on Vitex negundo L. – A Medicinally Important Plant

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ABSTRACT

Vitex negundo L. (Verbenaceae) is a hardy plant, flourishing mainly in the Indian subcontinent. All parts of the plant, from root to fruit, possess a multitude of phytochemical secondary metabolites which impart an unprecedented variety of medicinal uses to the plant. It is interesting to note that a single plant species finds use for treatment of a wide spectrum of health disorders in traditional and folk medicine; some of which have been experimentally validated. The plant is a component of a number of commercially available herbal formulations and has also shown potential as an effective bio-control agent. Employment of techniques such as cell and tissue culture would provide means of rapid propagation and conservation of the plant species and, from the point of view of phytochemistry, give scope for enhancement of the quality and quantity of the bioactive secondary metabolites occurring in the plant.

Key words: phytochemical; pharmacological; bio-control; herbal formulations; in vitro culture

Abbreviations:

BA: 6-Benzyl Amino Purine CH: Casein Hydrolysate GA₃: Gibberellic Acid IAA: Indole-3-Acetic Acid IBA: Indole-3-Butyric Acid Kn: Kinetin MS: Murashige and Skoog NAA: α-Naphthoxy Acetic Acid Vn: Vitex negundo

1. INTRODUCTION

1.1. Medicinal Plants

Morgenstern [1] defines ethnobotany as the study of the indigenous uses of plants and the relationship between people and plants. Folk medicines of almost all civilizations of the world abound in herbal remedies. Majority of the traditional medicines used in healthcare are obtained from plants [2]. In spite of several advancements in the field of synthetic drug chemistry and antibiotics, plants continue to be one of the major raw materials for drugs treating various ailments of humans. Clinical and pharmaceutical investigations have in fact elevated the status of medicinal plants by identifying the role of active principles present in them and elaborating on their mode of action in human and animal systems [3].

The world is gradually turning to herbal formulations which are known to be effective against a large repertoire of diseases and ailments. More importantly, they are not known to cause any notable derogatory effects [4]; and are readily availabile at affordable prices [5]. Prajapati et al. [6], however, add a note of caution stating that plant remedies are effective and without side-effects, provided they are selected properly and taken under proper medical supervision. The active component, most often a secondary metabolite, varies in quality and quantity for a given plant species growing in different locations. The market value of such plants depends on their active content rather than merely their luxuriant growth. Purohit and Vyas [7] reckon that close to 70,000 species of the plant kingdom have been used as herbal medicine at one time or other.

1.2. Vitex negundo Linn.

Vitex negundo Linn. (Verbenaceae) is a woody, aromatic shrub growing to a small tree. It commonly bears tri- or penta-foliate leaves on quadrangular branches, which give rise to bluish-purple coloured flowers in branched tomentose cymes. It thrives in humid places or along water courses in wastelands and mixed open forests and has been reported to occur in Afghanistan, India, Pakistan, Sri Lanka, Thailand, Malaysia, eastern Africa and

Madagascar. It is grown commercially as a crop in parts of Asia, Europe, North America and the West Indies [8]. Though *V. negundo* (will henceforth be referred to as Vn for sake of convenience) also finds use as a food crop (Facciola, 1990) and a source of timber [9], this review deals only with the medicinal importance and other related attributes of the plant.

2. Phytochemical Constituents

Higher plants are warehouses of assorted bioactive constituents or phytochemicals which find ample use in the pharmaceutical industry. Namdeo [10] states that about a quarter of all prescribed pharmaceuticals in advanced countries contain compounds that are directly or indirectly, derived from plants. Phytochemicals or secondary metabolites usually occur in complex mixtures that differ among plant organs and stages of development [11, 12]. Knowledge of the phytochemical constituents is very essential to enable investigation of the actual effectiveness of the plant in medicine. Table 1 gives the details of the different phytochemical constituents that have been reported from different parts of Vn.

PLANT PART	PHYTOCHEMICAL CONSTITUENTS	REFERENCE
Leaves hydroxy-3,6,7,3',4'-pentamethoxyflavone		[12]
	6'-p-hydroxybenzoyl mussaenosidic acid; 2'-p-hydroxybenzoyl mussaenosidic acid	[13, 14]
	5, 3'-dihydroxy-7,8,4'-trimethoxyflavanone; 5,3'-dihydroxy-6,7,4'-	[15]
	trimethoxyflavanone	
	viridiflorol; β-caryophyllene; sabinene; 4-terpineol; gamma-terpinene;	[16]
	caryophyllene oxide; 1-oceten-3-ol; globulol	
	betulinic acid [3β-hydroxylup-20-(29)-en-28-oic acid]; ursolic acid [2β -hydroxyurs-	[17]
	12-en-28-oic acid]; n-hentriacontanol; β-sitosterol; p-hydroxybenzoic acid	
	protocatechuic acid; oleanolic acid; flavonoids	[18]
	angusid; casticin; vitamin-C; nishindine; gluco-nonitol; p-hydroxybenzoic acid; sitosterol	[19]
Seeds	3β -acetoxyolean-12-en-27-oic acid; 2 α , 3 α -dihydroxyoleana-5,12-dien-28-oic acid;	[20, 21]
	2β,3α diacetoxyoleana-5,12-dien-28-oic acid; 2α, 3β-diacetoxy-18-hydroxyoleana- 5,12-dien-28-oic acid	
	vitedoin-A; vitedoin-B; a phenylnaphthalene-type lignan alkaloid, vitedoamine-A;	[22]
	five other lignan derivatives	
	6-hydroxy-4-(4-hydroxy-3- methoxy-phenyl)-3-hydroxymethyl-7-methoxy-3, 4- dihydro-2-naphthaldehyde	[23]
	β -sitosterol; p-hydroxybenzoic acid; 5-oxyisophthalic acid; n-tritriacontane, n-hentriacontane; n-pentatriacontane; n-nonacosane	[19]
Roots	2β , 3α -diacetoxyoleana-5,12-dien-28-oic acid; 2α , 3α -dihydroxyoleana-5,12-dien-28-	[24]
	oic acid; 2α , 3β -diacetoxy-18-hydroxyoleana-5, 12-dien-28-oic acid; vitexin and isovitexin	
	negundin-A; negundin-B; (+)-diasyringaresinol; (+)-lyoniresinol; vitrofolal-E and vitrofolal-F	[25]
	acetyl oleanolic acid; sitosterol; 3-formyl-4.5-dimethyl-8- oxo-5H-6,7-	[26]
	dihydronaphtho (2,3-b)furan	
Essential oil of	δ-guaiene; guaia-3,7-dienecaryophyllene epoxide; ethyl-hexadecenoate; α -selinene;	[27]
fresh leaves,	germacren-4-ol; caryophyllene epoxide; (E)-nerolidol; β -selinene; α -cedrene;	
flowers and dried fruits	germacrene D; hexadecanoic acid; p-cymene and valencene.	

Table 1 – Phytochemical constituents of different plant parts of V. negundo

2.1. Summary

Extensive biochemical analyses have resulted in the detection and isolation of a wide variety of the phytochemical constituents from different parts of the plant. Application of advanced spectroscopy tools such as NMR, EMR, FTIR along with X- ray crystallography studies would bring to light more such biologically active phytochemicals in different parts of the plant. Use of *in silico* tools to evaluate the efficacy of these phytochemical moieties as drugs would endow an added value to such a study.

3. Medicinal Importance

Herbal medicine, rather than merely curing a particular disease, aims at returning the body back to its natural state of health [28]. The phytochemical components of medicinal plants often act individually, additively or synergistically in improvement of health [29]. After having analyzed the various chemical components present in different parts of Vn, it is imperative that focus shifts to the medicinal applications of the plant. Myriad medicinal properties have been ascribed to Vn and the plant has also been extensively used in treatment of a plethora of ailments [6]. These properties have been categorized under three heads – traditional medicine, folk medicine and pharmacological evidence.

3.1. Traditional medicine

Traditional medicine mainly comprises of Indian Ayurveda, Arabic Unani medicine and traditional Chinese medicine. In Asia and Latin America, populations continue to use traditional medicine as a result of historical circumstances and cultural beliefs. Traditional medicine accounts for around 40% of all health care delivered in China. Up to 80% of the population in Africa uses traditional medicine to help meet their health care needs [30].

3.1.1. Ayurveda

The plant finds mention in the verses of the *Charaka Samhita* which is unarguably the most ancient and authoritative textbook of Indian Ayurveda. *Vn* has been designated as an anthelminthic (verse *Su:4-15*) and is prescribed as a vermifuge (verse *Vi:7-21*) in the exposition on the *Charaka Samhita* by Sharma [31].

Other Ayurvedic uses of *Vn* are described by Tirtha [32]. People sleep on pillows stuffed with *Vn* leaves to dispel catarrh and headache and smoke the leaves for relief. Crushed leaf poultice is applied to cure headaches, neck gland sores, tubercular neck swellings and sinusitis. Essential oil of the leaves is also effective in treatment of venereal diseases and other syphilitic skin disorders. A leaf decoction with *Piper nigrum* is used in catarrhal fever with heaviness of head and dull hearing. A tincture of the root-bark provides relief from irritability of bladder and rheumatism.

Jadhav and Bhutani [33] report the Ayurvedic use of *Vn* in dysmenorrhea. Patkar [34] refers to the formulations described in *Anubhoga Vaidya Bhaga*, a compendium of formulations in cosmetology, in outlining the use of *Vn* leaves along with those of *Azadirachta indica*, *Eclipta alba*, *Sphaeranthus indicus* and *Carum copticum* in a notable rejuvenation treatment known as *Kayakalpa*.

3.1.2. Unani medicine

Khare [19] outlines the applications of *Vn*, commonly known as *Nisinda* in Unani medicine. The seeds are administered internally with sugarcane vinegar for removal of swellings. Powdered seeds are used in spermatorrhoea and serve as an approdisiac when dispensed along with dry *Zingiber officinale* and milk.

3.1.3. Chinese medicine

The Chinese Pharmacopoeia prescribes the fruit of Vn in the treatment of reddened, painful, and puffy eyes; headache and arthritic joints [35].

3.2. Folk medicine

Folklore systems of medicine continue to serve a large segment of population, especially those in rural and tribal areas, regardless of the advent of modern medicine [36]. The entries regarding the multifarious applications of Vn in folk medicine have been grouped regionally to emphasize the ethnobotanical diversity and ubiquity of the plant; and the details have been laid out in Table 2 and 3.

S. NO.	STATE	REGION	LOCAL NAME	USED IN TREATMENT OF	REFERENCES
1 Andhra Pradesh			Tella Vaavili	Asthma, Cancer	[37]
		Puttaparthi		Used as bath for women in puerperal state and for new born children	Unpublished
				Jaundice	[38]
2	Assam		Pochatia	Urticaria, Cellulitis, Abcesses, Carbuncles, Eczema	[39]
				Liver disorders	[40]
2	II'm a shal Duada sh	Garwahl	Sambhaalu	Kwashiorkor	[4]
3 Himacha	Himachai Pradesh	Parvati valley	Bana	Wounds, Body ache	[41]
		Dharwad	Lakki, Karilakki	Toothache	[42]
4	Karnataka	Mysore	Bilenekki	Febrile, catarrhal and rheumatic afflictions	[4]
		Uttara Kanada	Nekki	Migraine	[43]
	Maharashtra	Konkan	Lingur	Rheumatism	[4]
5		Amravati	Samhalu	Encephalitis	[44]
5		Chota Nagpur	Nirgundi	Expectorant	[4]
		Satpuda	-	Joint pain	[36]
6	Orissa	Malkangiri	Languni	Jaundice	[45]
	Tamil Nadu	Southern parts	Notchi	Used as antidote for snake bite	[46]
		Madurai	-		[47]
7		Kancheepuram	-	Paspiratory disorders Favar Haadacha	[48]
		Salem and Tirucchirappalli	-	Respiratory disorders, rever, rieadache	[49]
8	Uttar Pradesh	Jaunsar-Bawar hills	Somi	Eye pain	[50]
		Moradabad	Mala	Used as refrigerant for cattle	[51]
		Uttaranchal	-	48 types of ailments	[2]

S. NO.	COUNTRY	REGION	LOCAL NAME	USED IN TREATMENT OF	REFERENCES
1	Bangladesh	Chittagong	-	Weakness, Headache, Vomiting, Malaria, Black fever	[52]
2	China	Guangdong	Buging'iab	Common cold, Flu and Cough	[53]
3	Nepal	Kali Gandaki	Simali	Sinusitis, Whooping cough	[54]
4	Pakistan	Buner	Marvandaey	Chest-pain, Backache Used as toothbrush	[55]
		Kot Manzaray Baba valley	-	Used as anti-allergenic agent	[56]
		Margallah hills	Nirgud	Gum and skin diseases	[9]
		Siran valley	Kalgari	Used as medicine for buffaloes in colic	[57]
5	Philippines		-	Cancer	[58]
6	Sri Lanka		Nilnikka	Eye disease, Toothache, Rheumatism Used as a tonic, carminative and vermifuge	[4]

Table 3 – Uses of V. negundo in folk medicine outside India

3.3. Pharmacological evidence

Demands of the scientific community have necessitated experimental evidence to further underline the medicinal importance of Vn described above. Taking cue from these traditional and folk systems of medicine, scientific studies have been designed and conducted in order to pharmacologically validate these claims.

3.3.1. Anti-inflammatory and analgesic activity

Yunos et al. [59] and Jana et al. [60] established anti-inflammatory properties of Vn extracts in acute and subacute inflammation. Anti-inflammatory and pain suppressing activities of fresh leaves of Vn are attributed to prostaglandin synthesis inhibition [61], antihistamine, membrane stabilising and antioxidant activities [62].

3.3.2. Effect on oxidative stress

Leaf extracts of Vn were determined to possess anti-oxidant potential by [63]. The extracts were useful in decreasing levels of superoxide dismutase, catalase and glutathione peroxidase in Freund's adjuvant induced arthritic-rats [64]. The extracts also possess the ability to combat oxidative stress by reducing lipid peroxidation owing to the presence of flavones, vitamin C and carotene [65]. Rooban et al. [66] evaluated the antioxidant and therapeutic potential of Vn flavonoids in modulating solenoid-induced cataract and found it to be effective.

3.3.3. Enzyme-inhibitory activity

Root extracts of Vn showed inhibitory activity against enzymes such as lipoxygenase and butyryl-cholinesterase [25]; α -chymotrypsin [67]; xanthine-oxidase [68] and tyrosinase [69]. Woradulayapinij et al. [70] reported the HIV type 1 reverse transcriptase inhibitory activity of the water extract of the aerial parts of Vn.

3.3.4. Effect on reproductive potential

The flavonoid rich fraction of seeds of Vn caused disruption of the latter stages of spermatogenesis in dogs [71] and interfered with male reproductive function in rats [72]. It must however be noted that these findings are in sharp contrast with the traditional use of Vn as approximately approximately and propounded its use in hormone replacement therapy.

3.3.5. Histomorphological and cytotoxic effects

Tandon and Gupta [74] studied the histomorphological effect of Vn extracts in rats and found the stomach tissue to be unaffected even by toxic doses; while dose-dependent changes were observed in the heart, liver and lung tissues. Cytotoxic effect of leaf extracts of Vn was tested and affirmed using COLO-320 tumour cells [75]. On one hand, Diaz et al. [76] found the chloroform extracts of Vn leaves to be toxic to a human cancer cell line panel while on the other; Yunos et al. [59] reported that Vn extracts were non-cytotoxic on mammary and genito-urinary cells of mice.

3.3.6. Drug potentiating ability

Administration of *Vn* extracts potentiated the effect of commonly used anti-inflammatory drugs such as ibuprofen and phenylbutazone [77]; analgesics such as meperidine, aspirin [78], morphine and pethidine; sedative-hypnotic drugs like pentobarbitone, diazepam [79] and chlorpromazine [80]; anti-convulsive agents such diphenylhydantoin and valporic acid [81].

3.3.7. Other attributes

In addition to the above mentioned activities Vn extracts have also been tested for a range of other systemic effects. Leaf extracts of Vn were found to possess hepato-protective activity against liver damage induced by d-galactosamine [82], commonly used tubercular drugs [83] and carbon tetrachloride [84, 85]. Villasenor and Lamadrid [86] have provided an account of the anti-hyperglycemic activity of Vn leaf extracts. Laxative activity of Vn leaf extracts was exhibited in rats by Adnaik et al. [87]. Methanolic root extracts of Vn showed antagonization of the lethal activity induced by venom of *Vipera russellii* and *Naja kaouthia* [88]. Immunomodulatory effect of Vn extracts has been reported by Ravishankar and Shukla [89].

3.4. Summary

Traditional medicine systems have also been developed throughout history by Asian, African, Arabic, Native American, Oceanic, Central and South American and other cultures. A comprehensive exploration in this regard could bring forth other medicinal applications of *Vn* hitherto unknown.

Ethnobotanical studies open many doors to knowledge that has rarely been cataloged. Such surveys have been conducted and reported from different regions of the Indian subcontinent. The listing of Vn in each of them has a novel account in terms of its medicinal use. Similar expeditions in other parts of India and countries where the plant is known to occur would bring to light the unknown aspects regarding this plant and other such plants that continue to remain obscure to the scientific community of the world at large. The lessons learnt from the traditional wisdom of the older generations combined with the modern scientific approach can provide the key to many of the unresolved issues of present-day medicine and open new vistas for the biotechnology industry.

Various other medicinal properties of Vn listed in the above sub-sections need to be established by systematic experimental studies before they can actually find their way to the market in the form of herbal-based medication for common ailments and afflictions. Such applications of Vn, on entering the pipeline of pharmacological research, could be taken up by the industry for Research and Development of drugs.

4. Commercial products

The pharmacological potential of Vn has been exploited effectively in formulating commercial products for treatment of health disorders. Table 4 provides details of such products available in the market which have supporting evidence in the form of scientific publications. Table 5 lists selected herbal products which contain Vn extracts, but do not have a backing in the form of readily available research literature.

MANUFACTURER	NAME OF PRODUCT	USED IN	
Himalaya Drug Co.,	Antiseptic Cream	Wounds, Burns, Fungal skin infections	
Bangalore, India	Dental Cream	Tooth ache, Bleeding gums	
	Himcolin Gel	Erectile dysfunction	
Surya Herbal Ltd., Noida,	Relieef Cream	Joint and Muscle pain, Stiff back	
India	Rheumanaad Tablet and	Rheumatic Pain, Sprain	
	Cream		
	Ostranil Gel	Osteoarthritis, Lumbago	
Ambica Research &	Amgesic Arthritis Tablets	Arthritis	
Development Pvt. Ltd, New			
Delhi, India			
IndSwift Ltd., Chandigarh,	Arthrill Capsules and	Arthritis, Joint pain, Frozen shoulder, Gout,	
India	Massage oil	Cervical spondylitis	

Table 4 - Commercial herbal formulations containing V. negundo

Table 5 - Commercial herba	l formulations	containing V	. negundo
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MANUFACTURER	NAME OF PRODUCT	USED IN	REFERENCE
Himalaya Drug Co., Bangalore	Acne-n-Pimple Cream	Acne and skin eruptions	[90]
	JointCare B cream	Rheumatic disorders	[91]
	Muscle and Joint Rub	Muscle strains, musculoskeletal disorders	[92]
	Pilex tablet and cream	Haemorrhoids (Piles)	[93, 94]
	Rumalaya gel and tablets	Inflammatory musculoskeletal disorders	[95-97]
	V-Gel	Vaginitis, Cervicitis	[98]
Hamdard Laboratories, New Delhi, India	Jigrine	Liver ailments	[99, 100]
Dey's Medical, Kolkata, India	Itone Eye Drops	Eye ailments	[101]

5. Biological activity

Plants are known to produce a variety of compounds which have evolved as defence compounds against microbes and herbivores [11]. The elaboration on the biochemically active ingredients and the medicinal properties of Vn elicits queries on the effect of the plant extracts on other biological organisms. Vn has shown promise as an effective bio-control agent. The extracts of Vn possess inhibitory, deterrent or lethal activity on biological agents that cause disease and damage to other organisms. Table 6 summarizes the effect of Vn on different pathogens and pests.

ACTIVITY	ACTION AGAINST	REFERENCE
Anti-bacterial	Escherichia coli, Klebsiella aerogenes, Proteus	[102]
	vulgaris, and Pseudomonas aerogenes (Bacteria)	
Anti-feedant	Spodoptera litura (Asian army-worm) Achoea	[17, 103]
	janata (Castor semi-looper)	
Anti-filarial	Brugia malayi (Microfilarial parasite)	[104, 105]
Anti-fungal	Alternaria alternata, Curvularia lunata	[106-108]
	Trichophyton mentagrophytes, Cryptococcus	
	neoformans	
	Aspergillus niger, Candida albicans	
Anti-larval	Cnaphalocrocis medinalis	[109]
	(Rice leaf-folder)	
Anti-viral	Plasmodium falciparum (Virus)	[110]
Insecticidal	Callosobruchus maculatus (Pulse beetle)	[111, 112]
	Phthorimaea operculella	[113]
	(Potato-tuber moth)	
	Sitotroga cerealella	[114]
	(Angoumois grain moth)	
	Aphis citricola (Spirea aphid), Aphis gossypii	[115]
	(Melon or Cotton aphid), <i>Myzus persicae</i> (Green peach aphid)	
Larvicidal	Anopheles subpictus,	[116]
	Culex tritaeniorhynchus (Mosquitoes)	
	Culex quinquefasciatus (Mosquito)	[117-121]
	Anopholos stophonsi (Mosquitoos)	[121]
	Anophetes stephensi (Mosquitoes)	[121]
	Plutella xylostella (Diamond-back moth)	[122]
Mosquito repellant	Culex tritaeniorhynchus (Mosquito)	[123]
Mosquito repellant	Aedes aegypti (Mosquito)	[124]

Table 6 - Activity of V. negundo extracts on biological pathogens and pests

6. In vitro culture

In vitro culture techniques offer viable means of mass multiplication and germplasm conservation of a multitude of plant species [125] and enhancement of the production of secondary metabolites. Due to poor viability of seeds of Vn [126] and slow rate of conventional propagation [127], tissue culture technology has assumed importance as an alternative method for rapid conservation and propagation of this economically important plant species. Additionally, production of plant secondary metabolites *de novo*, by *in vitro* cell culture methods, has assumed importance in the last two decades because the structural complexity of naturally occurring metabolites forms the basis for the chemical synthesis of novel and more potent analogues.

Nodal explants of *Vn* were found to be most effective for *in vitro* studies using Murashige and Skoog (MS) medium [128]. *In vitro* studies conducted to date have been preliminary; involving investigations on the response of explants to different phytohormones and growth supplements on MS medium.

6.1. Shoot initiation and proliferation

6-Benzyl Amino Purine (BA) in combination with α -Naphthoxy Acetic Acid (NAA) was found to be the most efficient in initiation of shoot primordial and formation of multiple shoots. Biotin and Casein Hydrolysate (CH)

[126]; sodium sulphate [129]; Gibberellic Acid (GA₃) [127]; Thidiaruzon [130, 131] and coconut water [131] gave positive results as growth supplements for shoot proliferation and multiple shoot formation. Sharief and Jagadishchandra [126] obtained shoots from stem callus using Kn and NAA in combination. Chandramu et al. [129] observed in vitro flowering with a combination treatment of BA and NAA. Handique [132] reported the use of Woody Plant medium but however found better response of explants in MS medium.

6.2. Root formation and elongation

Root formation and elongation was commonly observed with MS medium fortified with Indole-3-Butyric Acid (IBA) [129, 130, 132], NAA (Afroz et al., 2008), combination of NAA and Indole-3-Acetic Acid (IAA) [126, 131], combination of IAA and IBA [127, 133]. Usha et al. [133] made use of activated charcoal for rooting of *in vitro* plantlets. In contrast to other reports using half strength MS medium to induce rooting, Vadawale et al. [134] used full strength MS medium with IBA.

6.3. Summary

The prospects of rapid multiplication and propagation coupled with enhanced secondary metabolite production bring out with greater emphasis the need for establishment of standardized protocols for *in vitro* culture of medicinal plants, in this case *Vn*. Moreover, *in vitro* technology can potentially overcome common problems such as crop failure due to erratic weather conditions or mineral deficiencies in the soil. It is much simpler to manipulate and monitor the conditions essential for plant growth and development under laboratory conditions. Micropropagation also the advantage of rapid clonal multiplication of desired genotypes. Current approaches to *in vitro* propagation have shown encouraging results only utilizing MS medium as the substrate. Other nutrient media which have been shown to be useful for tissue culture of similar woody type of plants need to be tested, as also the role of other plant growth regulators and supplements.

Although India ranks among the top nations of the world medicinal plant exports, its export of phytochemical derivatives is insignificant in relation to developed countries of the world [135]. It is well-established that plants growing under stress conditions evolve to synthesize variants of commonly produced secondary metabolites. Cell culture provides means of bioconversion of low value compounds into high value products and enhancement of rate of production of these compounds. Application of established protocols using *Agrobacterium rhizogenes* (for hairy root cultures) and optimal elicitors needs to be investigated for enhancement of the yield of secondary metabolites in *Vn*. Additionally, *in vitro* methods can also be exploited to standardize the secondary metabolite content in a given amount of plant material which in turn would provided an added advantage in industrial applications of the plant.

7. CONCLUSION

Uniyal et al. [136] reiterates a popular local quote of the Bhangalis in the Western Himalayan region of India which translates as –"A man cannot die of disease in an area where *Vitex negundo*, *Adhatoda vasica* and *Acorus calamus* are found"; (provided that he knows how to use them). The plant holds great promise as a commonly available medicinal plant and it is indeed no surprise that the plant is referred to in the Indian traditional circles as *'sarvaroganivarini'* – the remedy for all diseases. Considerable amount of literature is available on various aspects of the plant – traditional to biochemical and ethnobotanical to pharmacological; however there many gaps which need to be filled by concurrent researchers in different disciplines. One must make the best use of the naturally available resources which provide valuable raw material for advanced research. Nature has many lessons to teach and the onus is on us to get attuned and grasp whatever is within our reach, before it is too late.

REFERENCES

[1] Morgenstern, K. (2001) 'What is ethnobotany?', 1st February, 2009.

[2] Kala, C.P., Farooquee, N.A. and Dhar, U. (2004) 'Prioritization of medicinal plants on the basis of available knowledge, existing practices and use value status in Uttaranchal, India', Biodiversity and Conservation. **13**, 453-469.

[3] Dutta, S.C. (1973) *Medicinal Plants*, National Council for Education Research and Training, New Delhi, 1973.

[4] Kirtikar, K.R. and Basu, B.D. (1984) *Indian Medicinal Plants*, Bishen Singh Mahendra Pal Singh, Dehradun, 1984.

[5] Sharma, A., Shanker, C., Tyagi, L.K., Singh, M. and Rao, C.V. (2008) 'Herbal medicine for market potential in India: An overview', Academic Journal of Plant Sciences. **1**, 26-36.

[6] Prajapati, D.S., Purohit, S.S., Sharma, A.K. and Kumar, T. (2004) *A Handbook of medicinal plants*, Agrobios India, Jodhpur, 2004.

[7] Purohit, S. and Vyas, S. (2004) 'Medicinal plant cultivation: A scientific approach', Agrobios, India. 624.

[8] de Padua, L.S., Bunyapraphatsara, N. and Lemmens, R.H.M.J. (1999) *Medicinal and Poisonous Plants*, Plant Resources of South East Asia, Backhuys Publishers, Leiden, 1999.

[9] Jabeen, A., Khan, M., Ahmad, M., Zafar, M. and Ahmad, F. (2009) 'Indigenous uses of economically important flora of Margallah Hills National Park, Islamabad, Pakistan', African Journal of Biotechnology. **8**, 763-784.

[10] Namdeo, A. (2007) 'Plant cell elicitation for production of secondary metabolites: A review', Pharmacognosy Reviews. **1**, 69-79.

[11] Wink, M. (2004) 'Phytochemical diversity of secondary metabolites', Encyclopedia of Plant & Crop Science, 915-919.

[12] Banerji, A., Chadha, M.S. and Malshet, V.G. (1969) 'Isolation of 5-hydroxy-3,6,7,3',4'-

pentamethoxyflavone from Vitex negundo', Phytochemistry. 8, 511-512.

[13] Sehgal, C.K., Taneja, S.C., Dhar, K.L. and Atal, C.K. (1982) '2'-p-hydroxybenzoyl mussaenosidic acid, a new iridoid glucoside from *Vitex negundo*', Phytochemistry. **21**, 363-366.

[14] Sehgal, C.K., Taneja, S.C., Dhar, K.L. and Atal, C.K. (1983) '6'-p-hydroxybenzoyl mussaenosidic acid, an iridoid glucoside from *Vitex negundo*', Phytochemistry. **22**, 1036-1038.

[15] Achari, B., Chowdhuri, U.S., Dutta, P.K. and Pakrashi, S.C. (1984) 'Two isomeric flavones from *Vitex negundo* ', Phytochemistry. **23**, 703-704.

[16] Singh, V., Dayal, R. and Bartley, J. (1999) 'Volatile constituents of Vitex negundo leaves', Planta medica. **65**, 580.

[17] Chandramu, C., Rao, D.M., Krupanandam, D.G.L. and Reddy, D.V. (2003) 'Isolation, characterization and biological activity of betulinic acid and ursolic acid from *Vitex negundo* L.', Phytotherapy Research. **17**, 129-134.

[18] Surveswaran, S., Cai, Y., Corke, H. and Sun, M. (2007) 'Systematic evaluation of natural phenolic antioxidants from 133 Indian medicinal plants', Food Chemistry. **102**, 938-953.

[19] Khare, C.P. (2004) Encyclopedia of Indian Medicinal Plants, Springer, Berlin, 2004.

[20] Chawla, A.S., Sharma, A.K. and Handa, S.S. (1992) 'Chemical investigation and anti-inflammatory activity of *Vitex negundo* seeds', Journal of Natural Products. **55**, 163-167.

[21] Chawla, A.S., Sharma, A.K., Handa, S.S. and Dhar, K.L. (1992) 'A lignan from *Vitex negundo* seeds', Phytochemistry. **31**, 4378-4379.

[22] Ono, M., Nishida, Y., Masuoka, C., Li, J., Okawa, M., Ikeda, T. and Nohara, T. (2004) 'Lignan derivatives and a norditerpene from the seeds of *Vitex negundo*', Journal of Natural Products. **67**, 2073-2075.

[23] Zheng, C.J., Tang, W.Z., Huang, B.K., Han, T., Zhang, Q.Y., Zhang, H. and Qin, L.P. (2009) 'Bioactivity-guided fractionation for analgesic properties and constituents of Vitex negundo L. seeds', Phytomedicine. **16**, 560-567.

[24] Srinivas, K.K., Rao, S.S., Rao, M.E.B. and Raju, M.B.V. (2001) 'Chemical constituents of the roots of *Vitex negundo*', Indian Journal of Pharmaceutical Sciences. **63**, 422-424.

[25] Azhar-Ul-Haq, Malik, A., Anis, I., Khan, S.B., Ahmed, E., Ahmed, Z., Nawaz, S.A. and I., C.M. (2004)
'Enzyme inhibiting lignans from *Vitex negundo*', Chemical and Pharmaceutical Bulletin. 52, 1269-1272.
[26] Vishnoi, S.P., Shoeb, A., Kapil, R.S. and Popli, S.P. (1983) 'A furanoeremophilane from *Vitex negundo*', Phytochemistry. 22, 597-598.

[27] Khokra, S., Prakash, O., Jain, S., Aneja, K. and Dhingra, Y. (2008) 'Essential oil composition and antibacterial studies of *Vitex negundo* Linn. extracts', Indian Journal of Pharmaceutical Sciences. **70**, 522-526.
[28] Srivastava, C. (2009) 'Phytochemistry and medicobotany of some medicinal plants used in treatment of arthritis', Medicinal Plants - International Journal of Phytomedicines and Related Industries. **1**, 27-32.

 [29] Schütz, K., Carle, R. and Schieber, A. (2006) 'Taraxacum—A review on its phytochemical and pharmacological profile', Journal of Ethnopharmacology. **107**, 313-323.

[30] WHO. WHO Traditional Medicine Strategy 2002–2005. World Health Organization 2002.

[31] Sharma, P.V. (2005) Caraka Samhita, Chaukhamba Orientalia, 2005.

[32] Tirtha, S. (1998) *The Ayurveda Encyclopedia*, Ayurveda Holistic Center Press, Bayville, New York, 1998.[33] Jadhav, A.N. and Bhutani, K.K. (2005) 'Ayurveda and gynecological disorders', Journal of

Ethnopharmacology. 97, 151-159.

[34] Patkar, K. (2008) 'Herbal cosmetics in ancient India', Indian Journal of Plastic Surgery. 41, 134-137.
[35] Liu, C., Tseng, A. and Yang, S. (2005) *Chinese herbal medicine: Modern applications of traditional formulas*, CRC Press, 2005.

[36] Kosalge, S.B. and Fursule, R.A. (2009) 'Investigation of ethnomedicinal claims of some plants used by tribals of Satpuda Hills in India', Journal of Ethnopharmacology. **121**, 456-461.

[37] Basavaraju, R., Vennel Raj, J. and Bhiravamurthy, P.V. (2009) 'Medicinal plant resources of Puttaparthi mandal: Taxonomic overview and need for conservation', Ethnobotanical Leaflets. **13**, 1382-1400.

[38] Purkayastha, J., Nath, S. and Islam, M. (2005) 'Ethnobotany of medicinal plants from Dibru-Saikhowa biosphere reserve of north-east India', Fitoterapia. **76**, 121-127.

[39] Saikia, A., Ryakala, V., Sharma, P., Goswami, P. and Bora, U. (2006) 'Ethnobotany of medicinal plants used by Assamese people for various skin ailments and cosmetics', Journal of Ethnopharmacology. **106**, 149-157.

[40] Kotoky, J. and Das, P. (2008) 'Medicinal plants used for liver diseases in some parts of Kamrup district of Assam, a north eastern state of India', Fitoterapia. **79**, 384-387.

[41] Sharma, P., Chauhan, N. and Lal, B. (2004) 'Observations on the traditional phytotherapy among the

inhabitants of Parvati valley in western Himalaya, India', Journal of Ethnopharmacology. 92, 167-176.

[42] Hebbar, S.S., Harsha, V.H., Shripathi, V. and Hegde, G.R. (2004) 'Ethnomedicine of Dharwad district in Karnataka, India—plants used in oral health care', Journal of Ethnopharmacology. **94**, 261-266.

[43] Bhandary, M.J., Chandrashekar, K.R. and Kaveriappa, K.M. (1995) 'Medical ethnobotany of the Siddis of Uttara Kannada district, Karnataka, India', Journal of Ethnopharmacology. **47**, 149-158.

[44] Jagtap, S.D., Deokule, S.S. and Bhosle, S.V. (2006) 'Some unique ethnomedicinal uses of plants used by

the Korku tribe of Amravati district of Maharashtra, India', Journal of Ethnopharmacology. 107, 463-469.

[45] Pattanaik, C., Reddy, S.C. and Murthy, M.S.R. (2008) 'An ethnobotanical survey of medicinal plants used by the Didayi tribe of Malkangiri district of Orissa, India', Fitoterapia. **79**, 67-71.

[46] Samy, R.P., Thwin, M.M., Gopalakrishnakone, P. and Ignacimuthu, S. (2008) 'Ethnobotanical survey of folk plants for the treatment of snakebites in Southern part of Tamilnadu, India', Journal of Ethnopharmacology. **115**, 302-312.

[47] Ignacimuthu, S., Ayyanar, M. and Sivaraman, K. (2006) 'Ethnobotanical investigations among tribes in Madurai District of Tamil Nadu (India)', Journal of Ethnobiology and Ethnomedicine. **2**, 25-31.

[48] Muthu, C., Ayyanar, M., Raja, N. and Ignacimuthu, S. (2006) 'Medicinal plants used by traditional healers in Kancheepuram District of Tamil Nadu, India', Journal of Ethnobiology and Ethnomedicine. 2, 43-52.
[49] Rajadurai, M., Vidhya, V., Ramya, M. and Bhaskar, A. (2009) 'Ethno-medicinal plants used by the traditional healers of Pachamalai hills, Tamilnadu, India', Ethnomedicine. 3, 39-41.

[50] Jain, S.P. and Puri, H.S. (1984) 'Ethnomedicinal plants of Jaunsar-Bawar hills, Uttar Pradesh, India', Journal of Ethnopharmacology. **12**, 213-222.

[51] Ali, Z.A. (1999) 'Folk veterinary medicine in Moradabad District (Uttar Pradesh), India', Fitoterapia. **70**, 340-347.

[52] Khan, N. and Rashid, A. (2006) 'A study on the indigenous medicinal plants and healing practices in Chittagong Hill tracts (Bangladesh)', African Journal of Traditional, Complementary and Alternative Medicines. 3, 37-47.

[53] Au, D.T., Wu, J., Jiang, Z., Chen, H., Lu, G. and Zhao, Z. (2008) 'Ethnobotanical study of medicinal plants used by Hakka in Guangdong, China', Journal of Ethnopharmacology. **117**, 41-50.

[54] Joshi, A.R. and Joshi, K. (2000) 'Indigenous knowledge and uses of medicinal plants by local communities of the Kali Gandaki watershed area, Nepal', Journal of Ethnopharmacology. **73**, 175-183.

[55] Hamayun, M. (2005) 'Ethnobotanical studies of some useful shrubs and trees of district Buner, NWFP, Pakistan', Ethnobotanical Leaflets. 9.

[56] Zabihullah, Q., Rashid, A. and Akhtar, N. (2006) 'Ethnobotanical survey of Kot Manzary Baba valley, Malakand Agency, Pakistan', Pakistan Journal of Plant Sciences. **12**, 115-121.

[57] Shah, G.M. and Khan, M.A. (2006) 'Common medicinal folk recipes of Siran Valley, Mansehra, Pakistan', Ethnobotanical Leaflets. **10**, 49-62.

[58] Graham, J.G., Quinn, M.L., Fabricant, D.S. and Farnsworth, N.R. (2000) 'Plants used against cancer–an extension of the work of Jonathan Hartwell', Journal of Ethnopharmacology. **73**, 347-377.

[59] Yunos, N.M., Mat Ali, R., Kean, O.B. and Abas, R. (2005) 'Cytotoxicity Evaluations on Vitex negundo Anti-inflammatory Extracts', Malaysian Journal of Science. **24**, 213-217.

[60] Jana, U., Chattopadhyay, R.N. and Shaw, B.P. (1999) 'Preliminary studies on anti-inflammatory activity of *Zingiber officinale* Rosc., *Vitex negundo* Linn. and *Tinospora cordifolia* (Willid) Miers in albino rats', Indian journal of pharmacology. **31**, 232-233.

[61] Telang, R.S., Chatterjee, S. and Varshneya, C. (1999) 'Studies on analgesic and anti-inflammatory activities of *Vitex negundo* Linn.', Indian journal of pharmacology. **31**, 363-366.

[62] Dharmasiri, M.G., Jayakody, J.R.A.C., Galhena, G., Liyanage, S.S.P. and Ratnasooriya, W.D. (2003) 'Antiinflammatory and analgesic activities of mature fresh leaves of *Vitex negundo*', Journal of Ethnopharmacology. **87**, 199-206.

[63] Tiwari, O.P. and Tripathi, Y.B. (2007) 'Antioxidant properties of different fractions of Vitex negundo Linn', Food Chemistry. **100**, 1170-1176.

[64] Devi, P.R., Kumari, S.K. and Kokilavani, C. (2007) 'Effect of *Vitex negundo* leaf extract on the free radicals scavengers in complete Freund's adjuvant induced arthritic rats', Indian Journal of Clinical Biochemistry. **22**, 143-147.

[65] Vishal, T. and Gupta, R.K. (2005) 'Effect of Vitex negundo on oxidative stress', Indian journal of pharmacology. **37**.

[66] Rooban, B., Lija, Y., Biju, P., Sasikala, V., Sahasranamam, V. and Abraham, A. (2009) 'Vitex negundo attenuates calpain activation and cataractogenesis in selenite models', Experimental Eye Research. 88, 575.
[67] Lodhi, A., Choudhary, I., Malik, A. and Ahmad, S. (2008) 'a-Chymotrypsin inhibition studies on the lignans from *Vitex negundo* Linn', Journal of Enzyme Inhibition and Medicinal Chemistry. 23, 400-405.
[68] Umamaheswari, M., AsokKumar, K., Somasundaram, A., Sivashanmugam, T., Subhadradevi, V. and Ravi, T.K. (2007) 'Xanthine oxidase inhibitory activity of some Indian medical plants', Journal of Ethnopharmacology. 109, 547-551.

[69] Azhar-Ul-Haq, Malik, A., Khan, M.T.H., Khan, S.B., Anwar-Ul-Haq, Ahmad, A. and Choudhary, M.I. (2006) 'Tyrosinase inhibitory lignans from the methanol extract of the roots of *Vitex negundo* Linn. and their structure–activity relationship', Phytomedicine. **13**, 255-260.

[70] Woradulayapinij, W., Soonthornchareonnon, N. and Wiwat, C. (2005) 'In vitro HIV type 1 reverse transcriptase inhibitory activities of Thai medicinal plants and Canna indica L. rhizomes', Journal of Ethnopharmacology. **101**, 84-89.

[71] Bhargava, S. (1989) 'Antiandrogenic effects of a flavonoid-rich fraction of *Vitex negundo* seeds: A histological and biochemical study in dogs', Journal of Ethnopharmacology. **27**, 327-339.

[72] Das, S., Parveen, S., Kundra, C.P. and Pereira, B.M.J. (2004) 'Reproduction in male rats is vulnerable to treatment with the flavonoid-rich seed extracts of *Vitex negundo*', Phytotherapy Research. **18**, 8-13.

[73] Hu, Y., Zhang, Q., Hou, T., Xin, H., Zheng, H., Rahman, K. and Qin, L. (2007) 'Estrogen-like activities in *Vitex* species from China determined by a cell based proliferation assay', Pharmazie. **62**, 872-875.

[74] Tandon, V. and Gupta, R.K. (2004) 'Histomorphological changes induced by Vitex negundo in albino rats', Indian journal of pharmacology. **36**, 176-177.

[75] Smit, H.F., Woerdenbag, H.J., Singh, R.H., Meulenbeld, G.J., Labadie, R.P. and Zwaving, J.H. (1995) 'Ayurvedic herbal drugs with possible cytostatic activity', Journal of Ethnopharmacology. **47**, 75-84.

[76] Diaz, F., Chavez, D., Lee, D., Mi, Q., Chai, H.B., Tan, G.T., Kardono, L.B.S., Riswan, S., Fairchild, C.R. and Wild, R. (2003) 'Cytotoxic flavone analogues of vitexicarpin, a constituent of the leaves of *Vitex negundo*', Journal of Natural Products. **66**, 865-867.

[77] Tandon, V.R. and Gupta, R.K. (2006) 'Anti-inflammatory Activity and Mechanism of Action of Vitex negundo Linn', International Journal of Pharmacology. **2**, 303-308.

[78] Gupta, R.K. and Tandon, V.R. (2005) 'An experimental evaluation of anticonvulsant activity of *Vitex negundo*', Indian Journal of Physiology and Pharmacology. **49**, 163-172.

[79] Gupta, M., Mazumder, U.K., Bhawal, S.R. and Swamy, S.M.K. (1997) 'CNS activity of petroleum ether extract of *Vitex negundo* Linn in mice', Indian Journal of Pharmaceutical Sciences. **59**, 240-245.

[80] Gupta, M., Mazumder, U.K. and Bhawal, S.R. (1999) 'CNS activity of *Vitex negundo* Linn. in mice', Indian Journal of Experimental Biology. **37**, 143-146.

[81] Tandon, V.R. and Gupta, R.K. (2005) 'An experimental evaluation of anticonvulsant activity of Vitexnegundo', Indian Journal of Physiology and Pharmacology. **49**, 199-205.

[82] Yang, L., Yen, K., Kiso, Y. and Hikino, H. (1987) 'Antihepatotoxic actions of formosan plant drugs', Journal of Ethnopharmacology. **19**, 103-110.

[83] Tandon, V.R., Khajuria, V., Kapoor, B., Kour, D. and Gupta, S. (2008) 'Hepatoprotective activity of Vitex negundo leaf extract against anti-tubercular drugs induced hepatotoxicity', Fitoterapia. **79**, 533-538.

[84] Tasduq, S.A., Kaiser, P.J., Gupta, B.D., Gupta, V.K. and Johri, R.K. (2008) 'Negundoside, an irridiod glycoside from leaves of Vitex negundo, protects human liver cells against calcium-mediated toxicity induced by carbon tetrachloride', World Journal of Gastroenterology. **14**, 3693-3709.

[85] Raj, P.V., Chandrasekhar, H.R., Vijayan, P., Dhanaraj, S.A., Rao, C.M., Rao, J.V. and Nitesh, K. (2008) '*In vitro* and *in vivo* hepatoprotective effect of *Vitex negundo* leaves', Pharmacology Online. **3**, 281-295.

[86] Villaseñor, I.M. and Lamadrid, M.R.A. (2006) 'Comparative anti-hyperglycemic potentials of medicinal plants', Journal of Ethnopharmacology. **104**, 129-131.

[87] Adnaik, R.S., Pai, P.T., Mule, S.N., Naikwade, N.S. and Magdum, C.S. (2008) 'Laxative Activity of *Vitex negundo* Linn. Leaves', Asian Journal of Experimental Sciences. **22**, 159-160.

[88] Alam, M.I. and Gomes, A. (2003) 'Snake venom neutralization by Indian medicinal plants (*Vitex negundo* and *Emblica officinalis*) root extracts', Journal of Ethnopharmacology. **86**, 75-80.

[89] Ravishankar, B. and Shukla, V. (2007) 'Indian Systems Of Medicine: A Brief Profile', African Journal of Traditional, Complementary and Alternative Medicines. **4**, 319-337.

[90] Ravichandran, G., Bharadwaj, V.S. and Kolhapure, S.A. (2004) 'Evaluation of efficacy and safety of Acne-N-Pimple cream in acne vulgaris ', The Antiseptic. **101**, 249-254.

[91] Venkataranganna, M.V., Gopumadhavan, S., Mitra, S.K. and Anturlikar, S.D. (2000) 'Anti-inflammatory activity of JointCare B, a polyherbal formulation ', Indian Drugs. **37**, 543-546.

[92] Rajanna, S. and Kolhapure, S.A. (2005) 'Evaluation of the efficacy and safety of "Muscle & Joint Rub" in the treatment of sprains, contusions and inflammatory musculoskeletal disorders ', The Antiseptic. **102**, 389-393.

[93] Sahu, M. and Srivastava, P. (2001) 'Clinical Study of Pilex Combination Therapy Vs Conventional Ayurvedic Therapy in the Management of Haemorrhoids ', The Indian Practitioner. 54, 799-805.
[94] Reddy, S.S., Nagabhushanam, M. and Rao, R.M. (1984) 'Role of Pilex Tablets and Ointment in the

[94] Reddy, S.S., Nagabhushanam, M. and Rao, R.M. (1984) Role of Pilex Tablets and Ointment in the Treatment of Piles and Fissures ', Probe. 23, 213-217.

[95] Dutta, N. and Bhattacharya, M.N. (1974) 'Clinical Trial of Rumalaya in Some Rheumatic Conditions ', Probe. **13**, 57-70.

[96] Sharma, A. and Kolhapure, S.A. (2005) 'Evaluation of the efficacy and safety of Rumalaya gel in the management of acute and chronic inflammatory musculoskeletal disorders: An open, prospective, noncomparative, phase III clinical trial ', Medicine Update. **12**, 39-45.

[97] Chakravaty, R.N. (1978) 'Therapeutic studies on Rumalaya in orthopaedic practice', Probe. 17, 323-326.
[98] Singh, I. (2001) 'Evaluation of V-Gel in Vaginitis and Cervicitis', The Antiseptic. 98, 6-9.

[99] Ahmad, A., Pillai, K.K., Najmi, A.K., Ahmad, S.J., Pal, S.N. and Balani, D.K. (2002) 'Evaluation of hepatoprotective potential of jigrine post-treatment against thioacetamide induced hepatic damage', Journal of Ethnopharmacology. **79**, 35-41.

[100] Najmi, A.K., Pillai, K.K., Pal, S.N. and Aqil, M. (2005) 'Free radical scavenging and hepatoprotective activity of jigrine against galactosamine induced hepatopathy in rats', Journal of Ethnopharmacology. **97**, 521-525.

[101] Mitra, A., Gupta, A. and Mukhopadhyay, D. (1986) 'An extended clinical study with herbal eye drops preparation', The Antiseptic. **83**, 567-569.

[102] Samy, R.P., Ignacimuthu, S. and Sen, A. (1998) 'Screening of 34 Indian medicinal plants for antibacterial properties', Journal of Ethnopharmacology. **62**, 173-182.

[103] Sahayaraj, K. (1998) 'Antifeedant effect of somne plant extracts on the Asian armyworm, *Spodoptera litura* (Fabricius)', Current Science. **74**, 523.

[104] Sahare, K.N., Anandhraman, V., Meshram, V.G., Meshram, S.U., Gajalakshmi, D., Goswami, K. and Reddy, M.V. (2008) 'In vitro effect of foue herbal plants on the motility of *Brugia malayi* microfilariae', Indian Journal of Medical Research. **127**, 467-471.

[105] Sahare, K.N., Anandhraman, V., Meshram, V.G., Meshram, S.U., Reddy, M.V., Tumane, P.M. and Goswami, K. (2008) 'Anti-microfilarial activity of methanoloic extract of *Vitex negundo* and *Aegle marmelos* and their phytochemical analysis', Indian Journal of Experimental Biology. **46**, 128-131.

[106] Guleria, S. and Kumar, A. (2006) 'Antifungal activity of some Himalayan medicinal plants using direct bioautography', Journal of Cell and Molecular Biology. **5**, 95-98.

[107] Sathiamoorthy, B., Gupta, P., Kumar, M., Chaturvedi, A.K., Shukla, P.K. and Maurya, R. (2007) 'New antifungal glycoside from *Vitex negundo*', Bioorganic and Medical Chemistry Letters. **17**, 239-242.

[108] Aswar, P.B., Khadabadi, S.S., Kuchekar, B.S., Rajurkar, R.M., Saboo, S.S. and Javarkar, R.D. (2009) '*In vitro* evaluation of anti-bacterial and anti-fungal activity of *Vitex nigundo* (Verbenaceae)', Ethnobotanical Leaflets. **13**, 962-967.

[109] Nathan, S.S., Kalaivani, K. and Murugan, K. (2006) 'Behavioural responses and changes in biology of rice leaffolder folowing treatment with a combination of bacterial toxins and botanical insecticides', Chemosphere. **64**, 1650-1658.

[110] Nguyen-Pouplin, J., Tran, H., Tran, H., Phan, T.A., Dolecek, C., Farrar, J., Tran, T.H., Caron, P., Bodo, B. and Grellier, P. (2007) 'Antimalarial and cytotoxic activities of ethnopharmacologically selected medicinal plants from South Vietnam', Journal of Ethnopharmacology. **109**, 417-427.

[111] Paneru, R.B. and Shivakoti, G.P. (2001) 'Use of botanicals for the management of pulse beetle (*Callosobruchus maculatus* F.) in lentil', Nepal Agriculture Research Journal. **4-5**, 27-30.

[112] Raja, N., Albert, S. and Ignacimuthu, S. (2000) 'Effect of solvent residues of *Vitex negundo* Linn. and *Cassia fistula* Linn. on pulse beetle, *Callosobruchus maculatus* Fab. and its larval parasitoid, *Dinarmus vagabundus* (Timberlake)', Indian Journal of Experimental Biology. **38**, 290.

[113] Das, G.P. (1995) 'Plants used in controlling the potato tuber moth, *Phthorimaea operculella* (Zeller)', Crop Protection. **14**, 631-636.

[114] Rajendran, S. and Sriranjini, V. (2008) 'Plant products as fumigants for stored-product insect control', Journal of Stored Products Research. 44, 126-135.

[115] En-shun, J., Ming, X., Yu-qing, L. and Yu-feng, W. (2009) 'Toxicity of *Vitex negundo* extract to aphids and its co-toxicity with imidacloprid', Chinese Journal of Applied Ecology. **20**, 686-690.

[116] Kamaraj, C., Rahuman, A. and Bagavan, A. (2008) 'Antifeedant and larvicidal effects of plant extracts against *Spodoptera litura* (F.), *Aedes aegypti* L. and *Culex quinquefasciatus* Say', Parasitology Research. **103**, 325-331.

[117] Karmegam, N., Sakthivadivel, M., Anuradha, V. and Daniel, T. (1997) 'Indigenous plant extracts as larvicidal agents against *Culex quinquefasciatus* Say.', Bioresource Technology. **59**, 137-140.

[118] Kannathasan, K., Senthilkumar, A., Chandrasekaran, M. and Venkatesalu, V. (2007) 'Differential larvicidal efficacy of four species of *Vitex* against *Culex quinquefasciatus* larvae', Parasitology Research. **101**, 1721-1723.

[119] Kannathasan, K., Senthilkumar, A., Venkatesalu, V. and Chandrasekaran, M. (2008) 'Larvicidal activity of fatty acid methyl esters of *Vitex* species against *Culex quinquefasciatus*', Parasitology Research. **103**, 999-1001.

[120] Rahuman, A., Bagavan, A., Kamaraj, C., Vadivelu, M., Zahir, A., Elango, G. and Pandiyan, G. (2009) 'Evaluation of indigenous plant extracts against larvae of Culex quinquefasciatus Say (Diptera: Culicidae)', Parasitology Research. **104**, 637-643.

[121] Pushpalatha, E. and Muthukrishnan, J. (1995) 'Larvicidal activity of a few plant extracts against *Culex quinquefasciatus* and *Anopheles stephensi*', Indian Journal of Malariology. **32**, 14-23.

[122] Yuan, L., Xue, M., Liu, Y. and Wang, H. (2006) 'Toxicity and oviposition-deterrence of Vitex negundo extracts to Plutella xylostella', Ying Yong Sheng Tai Xue Bao. **17**, 695-8.

[123] Karunamoorthi, K., Ramanujam, S. and Rathinasamy, R. (2008) 'Evaluation of leaf extracts of *Vitex negundo* L. (Family: Verbenaceae) against larvae of *Culex tritaeniorhynchus* and repellent activity on adult vector mosquitoes', Parasitology Research. **103**, 545-550.

[124] Hebbalkar, D.S., Hebbalkar, G.D., Sharma, R.N., Joshi, V.S. and Bhat, V.S. (1992) 'Mosquito repellant activity of oils from *Vitex negundo* Linn. leaves', Indian Journal of Medical Research. **95**, 200-203.

[125] Bajaj, Y. (1988) Biotechnology in agriculture and forestry Springer Verlag, Berlin, 1988.

[126] Sharief, M.U. and Jagadishchandra, K.S. (1992) 'Morphogenetic studies for callus cultures and plant regenration in *Vitex negundo* Linn.', Plant Science Research. **8**, 33-36.

[127] Sahoo, Y. and Chand, P. (1998) 'Micropropagation of Vitex negundo L., a woody aromatic medicinal shrub, through high-frequency axillary shoot proliferation', Plant Cell Reports. **18**, 301-307.

[128] Murashige, T. and Skoog, F. (1962) 'A revised medium for rapid growth and bioassays with tobacco tissue culture', Physiologia Plantarum. **15**, 473-497.

[129] Chandramu, C., Rao, D. and Reddy, V. (2003) 'High frequency induction of multiple shoots from nodal explants of *Vitex negundo* L. using sodium sulphate', Journal of Plant Biotechnology. **5**, 107-113.

[130] Ahmad, N. and Anis, M. (2007) 'Rapid clonal multiplication of a woody tree, *Vitex negundo* L. through axillary shoots proliferation', Agroforestry Systems. **71**, 195-200.

[131] Rani, D. and Nair, G. (2006) 'Effects of plant growth regulators on high frequency shoot multiplication and callus regeneration of an important Indian medicinal plant, nirgundi (Vitex negundo L.)', In Vitro Cellular & Developmental Biology-Plant. **42**, 69-73.

[132] Handique, P.J. (2007) 'Rapid *in vitro* propagation of *Vitex negundo* using nodal explants', ICFAI Journal of Biotechnology. **1**.

[133] Usha, P., Benjamin, S. and Raghu, K. (2007) 'An Efficient Micropropagation System for Vitex negando L., an Important Woody Aromatic Medicinal Plant, Through Shoot Tip Culture', Research Journal of Botany. **2**, 102-107.

[134] Vadawale, A., Barve, D. and Dave, A. (2006) 'In vitro flowering and rapid propagation of Vitex negundo L.--A medicinal plant', Indian Journal of Biotechnology. **5**, 112-116.

[135] Lambert, J., Srivastava, J. and Vietmeyer, N. (1997) *Medicinal plants: Rescuing a global heritage*, World Bank Publications, 1997.

[136] Uniyal, S., Singh, K., Jamwal, P. and Lal, B. (2006) 'Traditional use of medicinal plants among the tribal communities of Chhota Bhangal, Western Himalaya', Journal of Ethnobiology and Ethnomedicine. **2**, 14-21.